


REVIEW

Open Access



# Decoding microglial immunometabolism: a new frontier in Alzheimer's disease research

Eun Sun Jung<sup>1</sup>, Hayoung Choi<sup>1</sup> and Inhee Mook-Jung<sup>1,2,3\*</sup> 

## Abstract

Alzheimer's disease (AD) involves a dynamic interaction between neuroinflammation and metabolic dysregulation, where microglia play a central role. These immune cells undergo metabolic reprogramming in response to AD-related pathology, with key genes such as TREM2, APOE, and HIF-1 $\alpha$  orchestrating these processes. Microglial metabolism adapts to environmental stimuli, shifting between oxidative phosphorylation and glycolysis. Hexokinase-2 facilitates glycolytic flux, while AMPK acts as an energy sensor, coordinating lipid and glucose metabolism. TREM2 and APOE regulate microglial lipid homeostasis, influencing A $\beta$  clearance and immune responses. LPL and ABCA7, both associated with AD risk, modulate lipid processing and cholesterol transport, linking lipid metabolism to neurodegeneration. PPARG further supports lipid metabolism by regulating microglial inflammatory responses. Amino acid metabolism also contributes to microglial function. Indoleamine 2,3-dioxygenase controls the kynurenine pathway, producing neurotoxic metabolites linked to AD pathology. Additionally, glucose-6-phosphate dehydrogenase regulates the pentose phosphate pathway, maintaining redox balance and immune activation. Dysregulated glucose and lipid metabolism, influenced by genetic variants such as APOE4, impair microglial responses and exacerbate AD progression. Recent findings highlight the interplay between metabolic regulators like REV-ERB $\alpha$ , which modulates lipid metabolism and inflammation, and Syk, which influences immune responses and A $\beta$  clearance. These insights offer promising therapeutic targets, including strategies aimed at HIF-1 $\alpha$  modulation, which could restore microglial function depending on disease stage. By integrating metabolic, immune, and genetic factors, this review underscores the importance of microglial immunometabolism in AD. Targeting key metabolic pathways could provide novel therapeutic strategies for mitigating neuroinflammation and restoring microglial function, ultimately paving the way for innovative treatments in neurodegenerative diseases.

**Keywords** Microglia, Immunometabolism, Metabolic reprogramming, Neuroinflammation, APOE, TREM2, HIF, Hexokinase, A $\beta$ , Tau

## Background

Alzheimer's disease (AD) is a multifactorial and polygenic disorder resulting from complex interactions between various genetic and environmental factors [1]. The clinical heterogeneity of AD, characterized by diverse symptoms among patients [2–4], complicates therapeutic strategies. Additionally, AD is closely associated with metabolic syndrome, which includes hypertension, hyperlipidemia, obesity, and type 2 diabetes mellitus (T2DM) [5–8]. Given this complexity, the efficacy of monotherapies, such as A $\beta$ -targeting antibody

\*Correspondence:

Inhee Mook-Jung  
inhee@snu.ac.kr

<sup>1</sup> Convergence Dementia Research Center, Seoul National University College of Medicine, Seoul, South Korea

<sup>2</sup> Department of Biomedical Sciences, Seoul National University College of Medicine, Seoul, South Korea

<sup>3</sup> Korea Dementia Research Center, Seoul, South Korea



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>. The Creative Commons Public Domain Dedication waiver (<http://creativecommons.org/publicdomain/zero/1.0/>) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

treatments, remains limited, highlighting the growing need for combination therapy strategies [9–11].

Consistent with this perspective, genome-wide association studies (GWAS) have identified numerous genetic risk factors for late-onset Alzheimer's disease (LOAD), with a significant proportion being linked to microglial function, underscoring their central role in disease pathology [12–14]. These risk genes are involved in essential microglial processes, including A $\beta$  clearance, lipid metabolism, and inflammatory regulation, highlighting microglial dysfunction as a key driver of AD pathogenesis [14–16].

As the resident immune cells of the central nervous system (CNS), microglia play a crucial role in innate immune responses by rapidly detecting and responding to pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) [17–21]. Upon activation, microglia produce pro-inflammatory cytokines, and engage in phagocytic activity, making them key regulators of the pathological environment [22]. Consequently, their ability to clear A $\beta$  and modulate neuroinflammation is critical in AD progression, highlighting the regulation of neuroinflammatory pathways as a promising therapeutic approach for AD [23–26].

Importantly, microglial function is closely linked to metabolic processes. Microglial plasticity—the ability to functionally and phenotypically adapt to environmental stimuli [27, 28]—is tightly regulated by metabolic flexibility [29, 30]. This metabolic adaptability enables microglia to efficiently utilize glucose, amino acids, and lipids to sustain energy-intensive processes, such as pro-inflammatory cytokine production and phagocytosis, under both homeostatic and pathological conditions [29, 31–33].

Among the risk genes identified through GWAS, *TREM2* and *APOE* stand out as key regulators of multiple metabolic pathways, including glucose, lipid, and amino acid metabolism [34–37]. By influencing microglial energy balance, immune responses, and phagocytic activity, these genes further underscore their complex association with AD pathology [38–42].

Furthermore, recent multi-omics studies have revealed that distinct microglial phenotypes are strongly associated with specific metabolic pathways in the context of neurodegenerative diseases such as AD (Fig. 1, Table 1). These findings highlight the crucial role of microglial metabolic regulation in AD pathogenesis and suggest that targeting these pathways may provide novel therapeutic opportunities [43–45].

This review provides a comprehensive analysis of how key metabolic pathways—glucose, lipid, and amino acid metabolism—are altered in microglia under AD conditions and their functional implications in disease

progression. Each metabolic pathway is examined individually to elucidate its specific role in microglial dysfunction and its contribution to AD pathogenesis. Additionally, the impact of major AD risk genes, *TREM2* and *APOE*, on these metabolic processes is explored, emphasizing their regulatory influence on microglial energy balance and immune responses. Lastly, key molecular mediators involved in microglial metabolism are highlighted, offering insights into the intricate regulatory mechanisms that drive metabolic adaptations in AD.

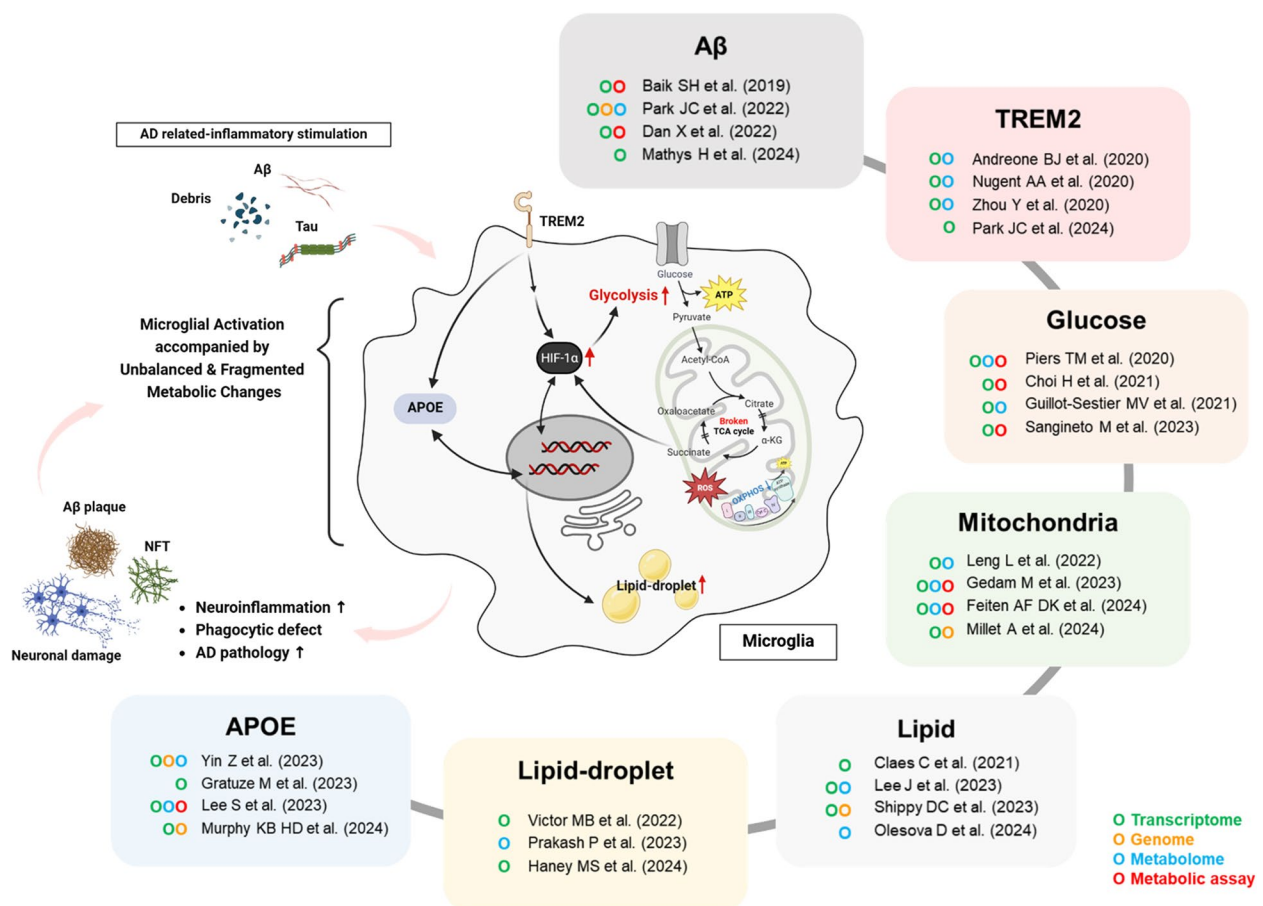
Microglial metabolic reprogramming is not merely a response to AD pathology [46–48] but actively drives disease progression, underscoring the critical role of immunometabolism in AD [49–51]. In AD, pathological stimuli such as amyloid-beta (A $\beta$ ) and tau aggregates induce shifts in glycolysis, oxidative phosphorylation (OXPHOS), and lipid metabolism [52–54] (Fig. 2). These changes are integral to microglial function, influencing their ability to clear toxic aggregates, modulate inflammation, and maintain homeostasis [49–51].

Glucose metabolism is vital for microglial function [51, 55, 56]. The brain, despite its small size, consumes about 20% of the body's energy, with glucose being the primary energy source. Microglia typically rely on oxidative phosphorylation for energy in homeostatic conditions but switch to glycolysis during inflammation, a shift that supports rapid energy production needed for immediate responses [57–59]. However, prolonged exposure to pathological stimuli such as A $\beta$  can lead to a state of microglial immune tolerance, in which both glycolysis and OXPHOS are downregulated, impairing microglial responsiveness and contributing to AD pathology [46, 60].

Lipid metabolism also plays a crucial role in microglial function [61–64]. A $\beta$ -driven lipid droplets (LDs) accumulation is linked to impaired phagocytosis and elevated reactive oxygen species (ROS) production, promoting neuroinflammation in AD [65, 66]. The involvement of a significant number of LOAD risk genes, including *TREM2* and *APOE*, in lipid metabolism underscores the close relationship between microglial lipid metabolism and AD pathology [34, 37, 67].

In addition to glucose and lipids, microglia exhibit metabolic flexibility by utilizing alternative energy sources, such as amino acids, particularly in circumstances where glucose availability is limited. This metabolic adaptation enables microglia to sustain their critical functions under varying nutritional conditions [30, 68]. However, dysregulation of amino acid metabolism can exacerbate microglial activation and neuroinflammation, particularly in microglia expressing the *APOE4* allele [69].

The metabolic pathways essential for microglial function are not independent but are tightly interconnected,



**Fig. 1** Major metabolic alterations and regulatory factors in AD microglia revealed by immuno-metabolomics approaches. Microglia function and phenotype change dynamically as AD pathogenesis progresses, accompanied by significant metabolic alterations. Initially, these cells demonstrate metabolic flexibility, but as the disease becomes chronic, microglia gradually lose their adaptive capacity. This reliance on biased and fragmented metabolic pathways eventually leads to insufficient energy and material supply, resulting in functional impairment and accelerated disease progression. The first notable metabolic shift in AD microglia involves glucose metabolism, characterized by excessive glucose uptake and increased dependence on non-aerobic glycolysis via HIF-1α pathway. This change subsequently leads to the inhibition and breakdown of mitochondrial energy metabolism. Furthermore, prolonged exposure to Aβ plaques, neurofibrillary tangles (NFTs), and excessive cell debris dramatically alters overall lipid metabolism, resulting in the accumulation of lipid droplets (LDs). Microglia with excessive LD accumulation, known as lipid-droplet-accumulating microglia (LDAMs), exhibit significantly reduced phagocytic ability and enhanced inflammatory properties. Recent studies have highlighted the crucial roles of APOE and TREM2, both high-risk genes for AD, in mediating these metabolic transitions in microglia. These genes are increasingly recognized as key players in modulating microglial function and metabolism in the context of AD. The advent of multi-omics approaches has accelerated the identification of candidate substances involved in microglial function and metabolic regulation pathways. This comprehensive analysis provides a deeper understanding of the complex interplay between immune response and metabolism in AD microglia

influencing and regulating each other in a coordinated manner. TREM2 and APOE act as key regulators of microglial metabolism, orchestrating both metabolic and immune functions [39, 70]. TREM2, a receptor expressed on microglia, influences their metabolic state, lipid handling, and inflammatory responses, and its loss-of-function variants are linked to an increased risk of AD [71]. Similarly, APOE, particularly the APOE4 isoform, alters microglial metabolism by disrupting multiple metabolic pathways [72–74], leading to impaired immune

responses and exacerbation of AD pathology. Furthermore, the intricate interplay between TREM2 and APOE adds another layer of complexity to the regulation of microglial metabolism and function.

Additionally, hypoxia-inducible factors (HIFs) and energy-sensing enzymes like adenosine monophosphate-activated protein kinase (AMPK) and hexokinase 2 (HK2) are crucial in regulating microglial metabolism [29, 75, 76]. HIFs drive metabolic reprogramming under hypoxic conditions [77], AMPK acts as an energy sensor

**Table 1** Summary of AD microglial omics and metabolism studies

Models	Sample types	Modulation	Key Findings	Screening Methods	References
AD animal model microglia	APP/PS1 (M,F; 16-18 M)	isolated adult mouse microglia	↑ Glycolysis in females vs. males	multiplexed gene expression analysis; LC/MS	Guillot-Sestier et al. (2021) [90]
	APP/PS1	isolated adult mouse microglia	TREM2 depletion	bulk RNA-seq; metabolomics; lipidomics	Feiten et al. (2024) [248]
	5XFAD (M,F; 3-7 M)	isolated adult mouse microglia	↑ LD accumulation, ↓ FFAs, ↑ TGs	Lipidomics	Prakash et al. (2024) [66]
	5XFAD (-; 2,6,9 M)	hippocampus	Altered glucose metabolism (transporters, glycolysis, OXPHOS)	scRNA-seq	Choi et al. (2021) [127]
	5XFAD (F; 9 M)	brain	↑ Glycolysis, mitochondrial repurposing, ↑ proton leak	scRNA-seq	Sangineto et al. (2023) [92]
	5XFAD (F; 7,5 M)	isolated adult mouse microglia	Aβ-induced acute inflammation & tolerance depend on glycolysis shift	bulk RNA-seq	Baik et al. (2019) [46]
	5XFAD (-; 12 M)	isolated adult mouse microglia	Glycolysis/H4K12la/PKM2 positive feedback loop exacerbates microglial dysfunction	genome-wide CUT&Tag analysis	Pan et al. (2022) [114]
	5XFAD (M,F; 1,3,6,8 M)	isolated adult mouse microglia	TREM2 required for full DAM activation (↑ phagocytosis & lipid metabolism)	scRNA-seq; ChIP	Keren-Shaul et al. (2017) [157]
	5XFAD (F; 8)	isolated adult mouse microglia, BMDM	TREM2 loss → AMPK-driven autophagy, ↓ anabolic & energetic metabolism	Microarray; EIS-MS/MS	Ulland et al. (2017) [35]
	5XFAD (M,F; 7,15 M)	brain	↓ Microglia-expressed protein abundance in Trem2-/-	snRNA-seq; proteomics	Zhou et al. (2020) [253]
	5XFAD:IPKR; 5XFAD:Elz <sup>A</sup> (M, F; 6 M); PS19IPKR; PS19ElzA (M,F; 8 M)	isolated adult mouse microglia; Microglia-specific ribosomal profiling (TRAP)	Glycolytic & lipid metabolism changes, toxic lipid secretion	snRNA-seq; TRAP seq	Flurry et al. (2024) [196]
	5XFAD; hAPOE (-; 2,24 M)	brain	TIM depletion in cellular energetics (TCA cycle, glycolysis, PPP); ↑ ROS detox pathways & amino acid metabolism	scRNA-seq/scATAC-seq multiome; interactome	Millet et al. (2024) [259]
	hAPOE KI (F; 3,12,24 M)	region-specific	APOE4 microglia: ↑ Aerobic glycolysis, ↑ Hif1α, altered lipid metabolism	snRNA-seq; spatial transcriptomics; metabolomics	Lee et al. (2023) [70]
	hAPOE KI; P301S:hAPOE KI; APP/PS1:hAPOE KI	brain; isolated adult mouse microglia	APOE4-ITGB8-TGFB as a negative regulator of MGNDR response; altered lipid species	scRNA-seq; ChIP-seq; lipidomics	Yin et al. (2023) [256]
	P301ShAPOE KI; P301S:APOE KO (male; 9 M)	hippocampus; BMDM	ApoE4 & tau-associated microglialosis → TREM2-independent lysosomal lipid accumulation	snRNA-seq (FANS)	Gratuze et al. (2023) [274]

Table 1 (continued)

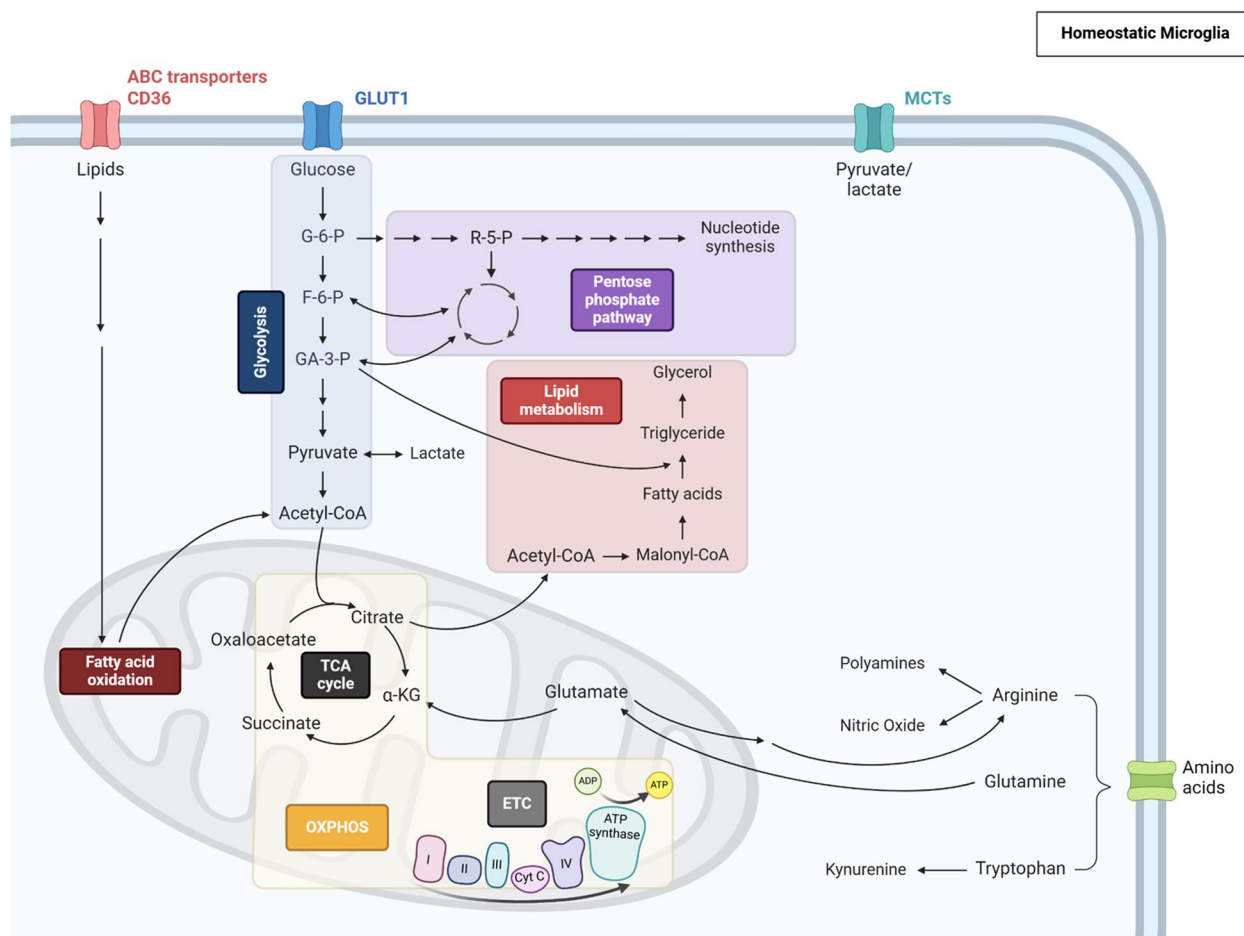
Models	Sample types	Modulation	Key Findings	Screening Methods	References
Human iPSC-derived microglia	APP NL-G-F (M/F; 9 M)	C3aRKO	C3aR + microglia: dysfunctional metabolism; C3aR1-null microglia resistant to hypoxia-induced lipid accumulation; C3aR ablation rescued lipid profiles & improved phagocytosis	bulk RNA-seq	Gedam et al. (2023) [309]
	APP <sup>5AA</sup> (M/F; 5-20 M)		Microglia with high intracellular Aβ → significant lipid & metabolite changes	bulk RNA-seq; LC/MS	Xia et al. (2022) [48]
	SHR24 (-; 4,6,8,10,12,14 M)	Pathological tau	↑ lipid production (protein fibrillization, membrane reorganization, inflammation); ↑ phospholipids, sphingolipids, & LD accumulation in microglia	Lipidomics	Olesova et al. (2024) [47]
	iPSC-derived microglia	TREM2-R47H variants	↓ OXPBOS & glycolysis; fails to switch to glycolysis after immune stimulation	Microarray; proteom array	Piers et al. (2020) [250]
	iPSC-derived microglia	PSEN1ΔE9, APPswe, and APOE4	APOE4 → defective glycolytic & mitochondrial metabolism	bulk RNA-seq	Konttinen et al. (2019) [257]
Human postmortem brain	iPSC-derived microglia	APOE3, APOE4; LD-high, LD-low	↑ LD accumulation, exacerbated by APOE4; LD spectra overlap with unsaturated TGs	bulk RNA-seq; ATAC-seq lipidomics	Haney et al. (2024) [260]
	iPSC-derived microglia and neuron (spheroid)	APOE3, APOE4	APOE4 → impaired lipid catabolism & lipid accumulation	bulk RNA-seq	Victor et al. (2022) [73]
	iPSC-derived microglia	APOE2, APOE3, APOE4, APOE KO	Altered cholesterol metabolism & lipid peroxidation	LCMS; RNA-seq	Guo et al. (2025) [283]
	AD-APOE4/4; AD-APOE3/3	frontal cortex	↑ LD accumulation, exacerbated by APOE4	snRNA-seq	Haney et al. (2024) [260]
	TREM2 variant carriers (R47H, R62H)	prefrontal cortex	TREM2-R47H → ↑ oxidative stress & lipid metabolism genes	snRNA-seq	Zhou et al. (2020) [253]
	AD-E4 patients; TREM2 variant carriers (R47H, R62H)	parietal cortex	APOE4 & tau → TREM2-independent lysosomal lipid accumulation	snRNA-seq	Gratuze et al. (2023) [274]
	AD-APOE3/4; AD-APOE3/3	brain	APOE4-ITGB8-TGFβ pathway inhibits MGN response; lipid species alterations	bulk RNA-seq	Yin et al. (2023) [256]
	ROSMAP	region-specific	↑ Glycogen-related gene expression → glial metabolic response may not be globally coordinated	snRNA-seq	Mathys et al. (2024) [357]
	AD-E3, E4 patients;	frozen superior frontal gyrus and fusiform gyrus	↑ Lipid accumulation	snRNA-seq	Haney et al. (2024) [260]

Table 1 (continued)

Models	Sample types	Modulation	Key Findings	Screening Methods	References
Xenotransplantation	TREM2-R47H xenografted microglia	iPSC-derived TREM2-R47H microglia transplantation	TREM2-R47H xMGs → ↓ LD accumulation & ↓ lipid metabolism genes (SPP1, APOE, CTSD)	scRNA-seq	Claes et al. (2021) [158]
	isolated microglia	iPSC-derived isogenic APOE microglia transplantation	Dysregulated lipid metabolism; ↓ CHCHD2 (mitochondrial migration gene)	bulk RNA-seq; ATAC-seq	Murphy et al. (2024) [261]

Abbreviations: Aβ amyloid beta, AD Alzheimer's disease, APOE apolipoprotein E, BMDM bone marrow-derived macrophages, Cε cholesterol ester, C3aR complement component 3a receptor, DAM disease-associated microglia, FFA free fatty acids, fAβ fibrillar amyloid beta, ICV intracerebroventricular, ISR integrated stress response, KJ knock-in, LD lipid droplet, LDLR low-density lipoprotein receptor, MgnD neurodegenerative microglia, OXPHOS oxidative phosphorylation, PPP pentose phosphate pathway, ROS reactive oxygen species, SPP1 secreted phosphoprotein 1, TCA tricarboxylic acid cycle, TG triacylglycerols, TIM terminally inflammatory microglia, TREM2 triggering receptor expressed on myeloid cells 2, xMGs xenografted microglia





**Fig. 2** An overview of major metabolic pathways. Cellular metabolism is a complex network of interconnected pathways that respond to internal and external signals to meet the cell's needs. These pathways function in a coordinated manner to produce essential components for cellular function and are subject to regulation by various signaling mechanisms. Here is an overview of the major metabolic pathways and their roles. Glycolysis is the process by which glucose is converted into pyruvate. Subsequently, pyruvate can further transform into lactate or be integrated into the tricarboxylic acid (TCA) cycle. Within the TCA cycle, pyruvate undergoes a series of reactions that produce NADH and FADH<sub>2</sub>, which the oxidative phosphorylation (OXPHOS) electron transport chain (ETC) system uses to generate ATP. Furthermore, glycolysis supplies intermediates for the pentose phosphate pathway (PPP), which produces ribose for nucleotides and amino acids. Lipid metabolism involves synthesizing fatty acids and lipid transportation, a process involving citrate derived from the TCA cycle. Moreover, it has been demonstrated that fatty acids can undergo oxidation, generating NADH and FADH<sub>2</sub>, which, in turn, promote ATP production through the OXPHOS. Amino acid metabolism also provides vital nutrients for the TCA cycle and plays a significant role in protein biosynthesis and adequate cellular activation. The intricate interconnection and regulation of these pathways by cellular signaling ensure metabolic activity aligns with the cell's requirements. This coordinated system enables cells to produce energy and essential molecules for various physiological processes efficiently

to maintain cellular energy homeostasis [78], and HK2 plays a pivotal role in glucose metabolism [79]. Dysregulation of these regulatory factors, which influence multiple metabolic pathways, may contribute to the chronic neuroinflammation and metabolic dysfunction observed in AD. Thus, in the later sections of this review, we will discuss how these key regulators of metabolic processes are modulated under AD conditions.

In summary, this review provides a comprehensive overview of metabolic pathways that regulate microglial function in AD. Starting with an exploration of energy

substrates like glucose, lipids, and amino acids, we then delve into molecular mediators such as TREM2, APOE, and other regulators that shape microglial immunometabolism. By integrating these findings, we aim to highlight therapeutic opportunities targeting these pathways.

## Main text

### Microglial energy substrates and metabolism in AD

#### Glucose metabolism

Glucose is the main source of energy for the cells in the CNS, including microglia [80]. These cells require

significant energy expenditure to perform critical functions such as surveillance and phagocytosis, relying on ATP, the primary cellular energy molecule, produced through various metabolic processes [81, 82]. Glycolysis and mitochondrial oxidative phosphorylation (OXPHOS) are the two major metabolic pathways that provide energy for microglia [51, 82]. Glycolysis involves a series of enzyme-catalyzed reactions that convert glucose to pyruvate or lactate. Following glycolysis, pyruvate is converted into acetyl coenzyme A (acetyl CoA) to enter the tricarboxylic acid (TCA) cycle for OXPHOS [83]. Although glycolysis produces ATP less efficiently than OXPHOS, it allows for rapid ATP production without requiring oxygen [84]. Typically, homeostatic microglia primarily rely on OXPHOS for ATP production, while inflammatory microglia are thought to switch to glycolysis through metabolic reprogramming to meet their energy needs [57–59]. This shift is advantageous for microglia as it allows them to respond quickly to danger signals [81, 85], a crucial capability in conditions such as AD.

Recent evidence indicates that microglia can rapidly switch their metabolism between glycolysis and OXPHOS depending on specific conditions like those present in AD [46, 86, 87]. For example, A $\beta$  has been shown to induce metabolic reprogramming in primary mouse microglia, shifting their preferred ATP production pathway from OXPHOS to glycolysis, a shift necessary for energy-intensive processes like phagocytosis and the production of pro-inflammatory cytokines [46, 86]. However, microglia exposed continuously to A $\beta$  may enter a state of tolerance, marked by a significant reduction in energy metabolism, encompassing both OXPHOS and glycolysis, leading to diminished responsiveness to external stimuli [46, 86]. This tolerance phase, characterized by reduced cytokine secretion and impaired phagocytosis, may contribute to the accumulation of A $\beta$  and the progression of AD pathology. Indeed, these characteristics of immune tolerance lend support to research findings suggesting that dying microglia with lost A $\beta$  phagocytic function may contribute to amyloid plaque growth, further exacerbating the disease process [88].

Multiple studies have reported alterations in glucose metabolism in microglia within various AD mouse models. For instance, both APP-PS1 [89–91] and 3xTg AD mice [92, 93] exhibit increased glycolysis and upregulation of glycolytic enzymes, although these effects diminish with age [92]. Similarly, microglia from 5xFAD mice display elevated levels of free NAD(P)H, indicating enhanced glycolytic activity [94]. However, it is essential to consider the broader metabolic landscape, as parallel pathways—such as the pentose phosphate pathway (PPP)—also contribute to NAD(P)H levels [95, 96]. In

response to immune activation by lipopolysaccharide (LPS) and interferon- $\gamma$  (IFN- $\gamma$ ), microglia shift their glucose metabolism toward both glycolysis and the PPP [97]. The PPP, which begins with glucose-6-phosphate, serves as a key metabolic pathway operating alongside glycolysis [96]. It provides essential cellular components such as NADPH, ribose-5-phosphate, and glycolytic intermediates. NADPH, in particular, is critical for regulating oxidative stress and maintaining reactive oxygen species (ROS) homeostasis [98, 99]. Its levels are primarily controlled by glucose-6-phosphate dehydrogenase (G6PD), the rate-limiting enzyme of the PPP [100, 101]. A delicate balance in G6PD activity is essential, as dysregulation can lead to oxidative stress-induced cellular damage [101, 102]. In microglia exposed to A $\beta$ , increased glucose consumption is accompanied by upregulated G6PD activity, leading to elevated mitochondrial ROS production and pro-inflammatory metabolic activation [103, 104]. NADPH also plays a significant role in lipid metabolism, as it is required for fatty acid synthesis [98, 105]. This underscores the intricate interconnectivity of metabolic pathways in microglia, demonstrating that glucose metabolism does not function in isolation but is closely coordinated with other metabolic processes [106, 107]. Understanding these interactions is crucial for elucidating microglial metabolic phenotypes in AD.

In addition to glucose, microglia can utilize lactate and pyruvate, key intermediates of glucose metabolism, as alternative energy sources [59, 107–109]. Upon exposure to inflammatory stimuli such as LPS or A $\beta$ , microglia undergo a metabolic shift from oxidative phosphorylation (OXPHOS) to glycolysis, leading to increased lactate production [46, 110]. While this shift initially supports an acute immune response, excessive lactate accumulation may also influence inflammation. Some studies suggest that exogenous lactate can mitigate LPS-induced inflammatory responses in both BV2 microglial cell lines and mice, indicating potential anti-inflammatory effects [111, 112]. However, other investigations have shown that lactate exposure impairs microglial phagocytic activity, reducing their ability to engulf and clear A $\beta$  aggregates [113]. Enhanced glycolysis in microglia can also lead to a self-reinforcing metabolic cycle, further amplifying glycolytic activity. This phenomenon is particularly evident in 5xFAD mice, where microglia surrounding A $\beta$  plaques exhibit a sustained glycolytic state [114]. A major contributing factor is lactate-induced histone lactylation, an epigenetic modification that enhances the expression of glycolysis-related genes, thereby reinforcing metabolic reprogramming [114, 115]. This positive-feedback loop disrupts microglial homeostasis, intensifies neuroinflammation, and ultimately accelerates AD progression. Interestingly, targeted inhibition of PKM2 (a key glycolytic



enzyme that promotes lactate production) in microglia of 5XFAD mice resulted in a reduction of amyloid plaques and improvements in cognitive function [114]. These findings suggest that glycolytic reprogramming and lactate metabolism play a crucial role in microglial-mediated neuroinflammation. However, further research is needed to fully elucidate the mechanistic link between lactate-driven epigenetic changes and microglial dysfunction in AD.

Microglia utilize glucose through various glucose transporters (GLUTs), with GLUT3 being the predominant isoform under normal physiological conditions. However, in inflammatory environments, GLUT1 expression is upregulated to facilitate increased glucose uptake and enhance glycolysis, thereby meeting the heightened energy demands associated with immune activation [116–118]. In AD, cerebral glucose metabolism declines significantly, a phenomenon strongly linked to cognitive impairment [119, 120]. Studies have reported reduced levels of GLUT1 in the brains of AD patients [121] and AD mouse models [122, 123]. This reduction in GLUT1 expression has been associated with exacerbated amyloid pathology and worsened cognitive function. Notably, in APP-PS1 model mice, GLUT1 levels in the hippocampus decline significantly only after extensive amyloid deposition [123], while cortical GLUT1 expression progressively decreases with aging in Tg2576 mice [124]. However, these studies did not specify the exact cellular contributors (e.g., neurons, microglia, or astrocytes) to the observed decrease in GLUT1 expression. While impaired glucose metabolism in neurons is well recognized as a driver of neurodegeneration [125, 126], the specific changes in microglial glucose metabolism and their contributions to AD progression remain less understood. Recent studies integrating 2-deoxy-2-[18F] fluoro-D-glucose positron emission tomography (FDG-PET) with single-cell RNA sequencing (scRNA-seq) have revealed a strong association between increased glucose uptake and metabolic reprogramming of microglia in the hippocampus of 5XFAD mice [127]. scRNA-seq analysis has identified distinct microglial subpopulations with unique glucose metabolism signatures, including a subset characterized by elevated GLUT1 expression, enhanced glycolysis, and reduced OXPHOS, which became more prevalent as AD pathology progressed [127]. These findings suggest that GLUT1 upregulation in microglia may serve as an adaptive response to sustain energy production in response to the disease. As AD progresses, glucose metabolism within the CNS becomes increasingly heterogeneous across different cell types [128–131]. Microglia exhibit diverse metabolic profiles depending on their activation states, which may also correlate with their proximity to amyloid plaques [94, 132]. A separate

FDG-PET study using an AD mouse model demonstrated that microglia exhibit higher glucose uptake than neurons and astrocytes [133]. Additionally, research utilizing TSPO-PET (18-kDa translocator protein PET) has established a positive correlation between microglial activation and glucose uptake in both AD and tauopathy mouse models, as well as in human patients [133]. These findings emphasize the importance of distinguishing between impaired neuronal glucose metabolism and microglial metabolic adaptation to accurately determine their respective contributions to AD pathology. Immune cells, including microglia, rely on glycolysis and mitochondrial metabolism to meet their energy and biosynthetic demands [134, 135].

Glycogen, a branched polymer of glucose, serves as a cellular energy reserve and a source of biosynthetic precursors [136]. While glycogen is primarily stored in the liver and muscles to provide energy when needed [137, 138], its role in the CNS has remained underexplored due to technical limitations in measuring glycogen levels [139]. This knowledge gap is particularly pronounced in microglia. However, recent technological advancements have enabled researchers to uncover novel insights into microglial glycogen metabolism [139].

Prostaglandin E2 (PGE2) is a bioactive lipid mediator that exerts its effects through four distinct G protein-coupled receptors (EP1–EP4), among which EP2 plays a critical role in inflammatory signaling and metabolic regulation [140, 141]. Recent studies in aged microglia have shown that activation of the PGE2-EP2 signaling pathway promotes glycogen synthesis, leading to glucose sequestration and reduced metabolic flux through glycolysis and mitochondrial respiration [142]. Interestingly, inhibition of the PGE2-EP2 pathway has been found to decrease glycogen accumulation, enhance glycolysis, and restore spatial memory and inflammatory responses in aged mice [142]. Glycogen accumulation has also been identified in specific microglial subpopulations [143, 144]. Among these, dark microglia (DM) represent a newly identified phenotype enriched under conditions of chronic stress, aging, and AD pathology [143, 145]. First described by Bisht et al., dark microglia exhibit ultrastructural features indicative of oxidative stress, including dense cytoplasm, nucleoplasmic condensation, dilated endoplasmic reticulum (ER), and mitochondrial abnormalities [143, 146]. Further studies by St-Pierre et al. identified glycogen granules as a unique feature distinguishing dark microglia from other microglial phenotypes [144]. Notably, glycogen-rich dark microglia are predominantly located near A $\beta$  plaques and dystrophic neurites in the hippocampus of APP-PS1 mice [144]. The presence of glycogen granules in these microglia suggests a potential metabolic shift toward glycolysis in response

to A $\beta$  plaque-induced stress. However, under pathological conditions, glycogen accumulation may also reflect impaired glucose metabolism, possibly due to decreased glucose availability. This metabolic dysfunction may compromise the microglial response to stressors such as A $\beta$ , ultimately influencing AD progression. Given the emerging role of glycogen metabolism in microglial function, further research is warranted to elucidate its relationship with glucose metabolism and its broader impact on AD pathogenesis.

A comprehensive understanding of glucose metabolism in AD requires an in-depth investigation of microglial metabolic profiles. However, current single-cell RNA sequencing (scRNA-seq) data are limited, as they are typically derived from a small number of mice and restricted to specific brain regions. This constraint hampers the ability to construct a holistic view of glucose metabolism across the entire brain in AD. To overcome this limitation, future scRNA-seq analyses should incorporate a broader dataset that accounts for key factors such as the spatial heterogeneity of microglia [147] and their proximity to amyloid plaques. These refinements will provide a more precise characterization of microglial glucose metabolism in AD and offer deeper insights into how metabolic alterations contribute to disease progression.

### **Lipid metabolism**

In addition to glucose, lipids serve as another crucial energy source influencing microglial activation. These lipids play essential roles in regulating membrane structure and cellular signaling within the brain. Microglia, equipped with diverse receptors, monitor the brain's microenvironment, with lipid rafts—sphingolipid- and cholesterol-rich microdomains in the plasma membrane—playing a significant role in processes such as receptor trafficking and signaling cascades [148, 149]. On the other hand, intracellular lipids, including those in mitochondria and signaling molecules, can modulate immune responses by affecting metabolic pathways [150]. Increasing attention is being directed toward the role of microglia-mediated lipid and lipoprotein metabolism in the brain and its implications for systemic and neuroinflammatory diseases [62]. The rapid growth of scRNA-seq studies has revealed a wide range of microglial phenotypes, far beyond the classical M1/M2 types [151–153]. These studies have highlighted changes in gene expression related to lipid and lipoprotein metabolism and their association with various microglial phenotypes [154]. Moreover, accumulating genetic and biological evidence underscores the significant role of lipid metabolism in AD. Over a century ago, Alois Alzheimer described 'lipid granules' in AD brains [155], raising early suspicions about the potential role of disrupted

lipid metabolism in the disease. These suspicions have since been reinforced by genome-wide association studies (GWAS), which have identified several lipid-related genes, including APOE, APOJ, and ABCA7, as strong AD risk factors [13, 15, 156]. Recent comprehensive single-cell transcriptomic analyses, including scRNA-seq and single-nucleus RNA sequencing (snRNA-seq), have further defined the transcriptional identities of microglia with disease-specific states [157–160].

For instance, Keren-Shaul et al. identified a unique population of microglia, termed disease-associated microglia (DAM), in AD mouse model [157]. Key genes upregulated in DAM, such as *APOE*, *TREM2*, *LPL*, and *CST7* are involved in lipid metabolism and phagocytosis [157, 161–163]. This transcriptional profile suggests that, in the later stages of AD, microglia may shift toward lipid metabolism as a primary energy source for A $\beta$  clearance. However, findings from DAM in mouse models may not fully capture the complexity of microglial states in human AD [164–166]. Recent snRNA-seq analyses by Sun et al. have expanded this understanding by revealing a greater diversity of microglial transcriptional states in human AD brains compared to the DAM signatures observed in AD mouse models [160]. These microglial substates were functionally distinct, characterized by features such as ribosome biogenesis, inflammatory responses, and lipid processing. A distinct microglial state linked to cholesterol and lipid homeostasis—characterized by genes such as *PPARG*, *APOE*, *ABCA1*, and *TREM2*—was significantly enriched in AD [160]. This microglial state showed a strong correlation with amyloid plaques, tau tangles, Braak staging, and cognitive decline. Furthermore, state- and stage-specific differential analyses revealed that inflammatory responses precede lipid regulation in microglia during disease progression [160]. These findings underscore the strong link between inflammation and lipid metabolism, emphasizing the critical role of microglia in AD pathogenesis.

Marschallinger et al. discovered a novel microglia subpopulation, lipid-droplet-accumulating microglia (LDAM) in the aging brain [65]. These microglia exhibit defective phagocytosis, excessive reactive oxygen species (ROS) production, and elevated pro-inflammatory cytokine secretion [65, 167]. LDAM exhibit a distinct transcriptome signature, setting them apart from previously described microglia states observed in aging and neurodegeneration, such as DAM and neurodegenerative microglia (MGnD). LDAM were also identified near amyloid plaques in 5XFAD mice [66] and in chimeric human–mouse AD models [158]. Similar cells have been observed in hippocampal tissue from AD patients [66], suggesting that LDAM may play a significant role in the progression of AD and other neurodegenerative

disorders. These findings highlight how lipid dysregulation in microglia may contribute to neurodegeneration.

Lipid metabolism, encompassing processes such as lipid uptake, lipolysis, fatty acid beta oxidation (FAO), lipogenesis, and fatty acid synthesis, provides essential energy for microglial activation [62]. Microglia express a wide range of genes related to lipid metabolism, including those involved in FAO [29, 168], a process crucial for their metabolic function. During FAO, fatty acids (FAs) are liberated from lipid droplets (LDs) and transported into mitochondria, where they fuel OXPHOS, generating the energy needed for cellular activities [169, 170]. This pathway becomes particularly important when glucose availability is limited, as demonstrated in studies on macrophages, which share many functional similarities with microglia [171, 172]. In energy-stressed environments, FAs serve as a vital alternative energy source, enabling microglia to maintain their essential functions in the CNS [173, 174]. Changes in FAO can affect microglial energy production and phenotype, potentially influencing their response to AD pathology [175, 176].

Lipoprotein lipase (LPL), a DAM-associated gene and an identified AD risk factor [157, 177], plays a crucial role in lipid metabolism in microglia, regulating lipid droplet accumulation and FAO [178, 179]. Loss of LPL in microglia leads to increased pro-inflammatory lipid profiles, impaired lipid uptake, and reduced FAO [178], contributing to neuroinflammation and microglial dysfunction. In AD, this impaired LPL function disrupts lipid homeostasis, facilitating the accumulation of neurofibrillary tangles and amyloid plaques [62, 177, 180]. Notably, in response to A $\beta$  exposure, microglia upregulate LPL expression, which enhances their phagocytosis of A $\beta$  [75, 181]. Additionally, in an AD mouse model, pharmacological inhibition of hexokinase 2 (HK2), a key glycolytic enzyme, increased LPL expression, which in turn regulated lipid metabolism and was associated with reduced amyloid load and improved cognitive function [75]. Similarly, ABCA7, another LOAD risk gene involved in lipid transport, is critical for maintaining phospholipid and cholesterol homeostasis [182, 183]. Genetic studies have linked ABCA7 loss-of-function variants to an increased risk of AD [184–187], yet the precise mechanisms underlying this association remain incompletely understood. Recent evidence suggests that ABCA7 is involved in the crosstalk between lipid metabolism and neuroinflammation. Metabolomic network analysis identified glycerophospholipid metabolism, linoleic acid metabolism, and  $\alpha$ -linolenic acid metabolism as key pathways affected by ABCA7 haploinsufficiency following acute inflammatory stimulation [188]. This was accompanied by increased levels of eicosapentaenoic acid, oleic acid, and palmitic acid, further supporting ABCA7's role in coordinating

lipid metabolism and immune responses [189]. In addition, a metabolome-wide association study (MWAS) has identified a link between ABCA7 and peripheral blood lactosylceramide (LacCers) levels, suggesting a potential role in sphingolipid metabolism [189]. Further analysis in ABCA7 knockout mice revealed decreased LacCer levels in the brain, while overall sphingolipids, ceramides, and hexosylceramides were increased, indicating that ABCA7 may be required for proper processing of sphingolipids into LacCer species [189]. These findings suggest that ABCA7 deficiency alters ceramide metabolism, potentially contributing to microglial lipid imbalance and neuroinflammation.

ABCA7-deficient microglia exhibit reduced A $\beta$  phagocytic clearance, which aligns with the increased A $\beta$  accumulation observed in the brains of ABCA7-deficient mice [190, 191]. Additionally, the rare ABCA7 p.A696S variant has been associated with exacerbated amyloid burden and plaque-associated neuritic dystrophy in 5xFAD mice, along with reduced amyloid-induced microglial activation and altered brain lipid profiles, including increased lysophosphatidylethanolamine (LysoPE) levels [192]. Transcriptomic analysis of ABCA7-A696S knock-in 5XFAD mice further revealed increased expression of DAM genes including *APOE*, *TREM2*, and *AXL* [192]. These findings suggest that ABCA7 dysfunction may disrupt microglial lipid metabolism and immune regulation, thereby contributing to AD pathogenesis by impairing A $\beta$  clearance and promoting neuroinflammation.

Microglia accumulate LDs when exposed to A $\beta$ , particularly near amyloid plaques in AD patients and mouse models [66, 193]. This lipid accumulation impairs microglial A $\beta$  clearance [66, 193]. Lipidomic analyses have shown a significant decrease in free fatty acids (FFAs) and an increase in triacylglycerols (TAGs) as the key metabolic transition underlying LD formation in microglia exposed to A $\beta$  [66]. This shift in lipid composition is mediated by DGAT2, which converts FFA to TAG [194, 195]. Increased levels of DGAT2 are found in microglia from AD brains [66]. Interestingly, the inhibition of DGAT2 has been shown to improve microglial uptake of A $\beta$ , suggesting a potential therapeutic strategy for reducing amyloid burden in AD [66]. Shippy et al. recently conducted a comprehensive analysis of lipid metabolism-related gene expression changes in microglia under AD and neuroinflammatory conditions [176]. Their microarray analysis of 5XFAD mice revealed numerous differentially expressed genes (DEGs) related to lipid metabolism compared to wild-type mice. Functional enrichment and network analyses identified several biological processes and molecular functions associated with these DEGs, including cholesterol homeostasis (e.g., *HMGCR*, *ANGPTL3*, *APOE*, *INSIG1*, *LPL*), fatty acid metabolic/

biosynthetic process (e.g., *ANGPTL3*, *ACADS*, *ACACA*, *FABP3*), and triglyceride metabolism (e.g., *HMGCR*, *ANGPTL3*, *APOC4*, *APOE*, *LPL*) [176]. Furthermore, recent study has demonstrated that reducing LD accumulation in microglia restores A $\beta$  phagocytosis and leads to a reduction in amyloid plaque deposition, highlighting the therapeutic potential of targeting microglial lipid metabolism in AD [193].

Building on these findings, recent studies have further elucidated the molecular mechanisms that link microglial lipid metabolism to AD pathology, identifying key signaling pathways involved in this process. Notably, two recent studies have highlighted the roles of the integrated stress response (ISR) [196] and REV-ERB $\alpha$  signaling [197] in mediating microglial lipid dysregulation and dysfunction. ISR activation in microglia, observed in both AD patients and mouse models, promotes the secretion of toxic lipids and impairs neuronal survival. This response exacerbates synapse loss and tau pathology, positioning ISR-activated microglia as a critical neurodegenerative phenotype [196]. Similarly, REV-ERB $\alpha$ , known for regulating lipid metabolism in the periphery, also modulates lipid metabolism and inflammatory responses in microglia within the CNS. Microglial REV-ERB $\alpha$  deletion leads to excessive lipid droplet accumulation, impaired tau clearance, and exacerbated neuroinflammation in tauopathy model [197]. This effect was observed specifically in male mice, highlighting the potential sex-dependent role of REV-ERB $\alpha$  in microglial lipid metabolism and tau pathology [197]. Together, these recent findings emphasize how specific signaling pathways mediate the link between lipid metabolism dysregulation and microglial dysfunction, offering potential avenues for therapeutic intervention in AD.

Emerging evidence suggests that, beyond the role of lipid metabolism in microglial dysfunction, pathogenic tau directly disrupts lipid metabolism, further exacerbating neurodegeneration. Olesova et al. demonstrated significant alterations in mitochondrial-associated lipid metabolism during various stages of tau pathology in a transgenic rat model (SHR-24) expressing human truncated tau protein [47]. Their analysis revealed that tau accumulation disrupts metabolic pathways, including FAO, TCA cycle, and creatine metabolism, contributing to mitochondrial dysfunction. In vitro studies further showed that truncated tau increases LD formation in microglial mitochondria, reducing ATP production and exacerbating oxidative stress [47]. Similarly, in PS19 mice expressing human P301S mutant tau, microglia enriched in LDs displayed elevated oxidative stress and an inflammatory phenotype, impairing tau clearance and driving neurodegeneration [198]. These findings highlight how lipid dysregulation and LD accumulation disrupt

mitochondrial function and microglial activity, creating a vicious cycle that amplifies both amyloid and tau pathologies.

Additionally, tau-induced changes in cellular membrane fluidity and lipid order further disrupt microglial function [47]. Alterations in lipid composition can significantly affect receptor expression and receptor-mediated signaling pathways, modulating cell polarization and activity [61, 149, 199, 200]. In microglia, tau has been shown to decrease membrane lipid order, shifting it toward a more liquid phase, potentially contributing to energy imbalance and impaired microglial responses [47]. These findings indicate that tau pathology may disrupt lipid metabolism and energy homeostasis in microglia, potentially contributing to neurodegenerative processes.

A recent study by Li et al. identified a potential mechanism underlying excessive LD accumulation in microglia in tauopathy models [198]. The researchers found that tauopathy neurons release excess unsaturated lipids, which are then taken up by nearby microglia, leading to significant LD accumulation in these cells [198]. Notably, LD accumulation in microglia appears to be associated with the exacerbation of tau pathology [201]. Importantly, the extent of LD buildup in microglia has been found to correlate with hippocampal neuron loss [198], suggesting a potential association between lipid dysregulation and neurodegeneration in tauopathies.

#### **Amino acids metabolism**

Microglia demonstrate remarkable metabolic flexibility in their energy acquisition. When glucose availability is limited, they can efficiently shift their metabolism to utilize amino acids as an alternative fuel, highlighting their bioenergetic adaptability [30]. Beyond their role in protein synthesis, amino acids can be catabolized through various pathways to generate ATP, providing an alternative energy source when needed [202]. For instance, glutamine act as a crucial bioenergetic substrate for microglia under hypoglycemic conditions [30]. Glutamine, the most abundant free amino acid in the brain, is converted into glutamate by glutaminase and then further transformed into  $\alpha$ -ketoglutarate by glutamate dehydrogenase, enabling it to enter the TCA cycle to support mitochondrial metabolism for the production of energy [203]. However, imbalances in glutamine metabolism during microglial inflammatory activation can potentially lead to detrimental outcomes [204–206]. Glutamine metabolism, particularly through the enzyme glutaminase (GLS), appears to regulate microglial activation and pro-inflammatory responses [207]. Abnormal elevation of the GLS isoform glutaminase C (GAC), the rate-limiting enzyme in glutaminolysis, has been observed in activated microglia and early AD mouse brain tissues [208].



Gao et al. demonstrated that GAC upregulation in microglia enhances the release of pro-exosome, potentially exacerbating neuroinflammation in AD [208]. Therefore, dysregulation of glutamine utilization may contribute to chronic neuroinflammation and microglial dysfunction in AD pathogenesis.

The APOE4 allele is well known as the greatest genetic risk factor for LOAD. Microglia expressing APOE4 exhibit exaggerated immune responses and a diminished capacity to clear A $\beta$  compared to microglia expressing APOE2 or APOE3 [209, 210]. These functional differences underscore the impact of APOE genotype on microglial behavior in the context of AD. Notably, a significant increase in GLS activity was observed in hippocampal microglia from APOE4 knock-in mice compared to age-matched wild-type controls [69]. Chronic administration of JHU-083, a glutaminase antagonist, improved cognition and attenuated the rise of GLS activity in APOE4 knock-in mice [69]. Collectively, these studies indicate that while glutamine is a vital metabolic substrate for microglia, its excessive availability or altered metabolism during pro-inflammatory responses can potentially contribute to neuroinflammation and neurotoxicity.

Alongside glutamine, other amino acids, including tryptophan and arginine, also play crucial roles in microglial metabolism and immune responses [211, 212]. Tryptophan serves as a precursor for neuroactive compounds and influences microglial function, contributing to either neurotoxic or neuroprotective effects [213]. The majority of tryptophan is metabolized via the kynurenine pathway, and alterations in this metabolic pathway have been linked to various human diseases, including psychiatric disorders, autoimmunity, neurodegeneration, and AD [51, 214–217]. The rate-limiting enzyme in the kynurenine pathway, indoleamine 2,3-dioxygenase (IDO), is upregulated in the AD brain and promotes the conversion of tryptophan into quinolinic acid (QA) [215, 218]. This pathway is particularly active in microglia exposed to A $\beta$ , leading to an increase in QA production. Elevated IDO levels in AD are associated with neurofibrillary tangles and A $\beta$  plaques, suggesting a potential role of tryptophan metabolism in AD pathology [215, 218, 219]. Additionally, QA, a key metabolite of tryptophan degradation, has been implicated in neurotoxicity in various inflammatory brain disorders, including AD [215, 220, 221]. During neuroinflammation, microglial activation promotes the shift in kynurenine metabolism towards increased QA production [222, 223]. QA is secreted from microglia and acts as an NMDA receptor agonist, inducing neurotoxicity [224]. Furthermore, studies have shown that QA treatment induces tau phosphorylation in neurons [225].

Arginine is metabolized through two competing pathways: one via nitric oxide synthase (iNOS) to generate nitric oxide (NO) and citrulline, and the other via arginase to produce urea and ornithine [226]. iNOS is a marker of the pro-inflammatory M1 microglial phenotype, while arginase-1 is associated with the anti-inflammatory M2 phenotype [227–229]. NO, synthesized through iNOS, inhibits Complex IV of the mitochondrial electron transport chain, disrupting mitochondrial function and shifting microglial metabolism toward glycolysis [230]. NO produced by iNOS, upregulated in microglia upon A $\beta$  exposure, has been shown to induce neurotoxicity [231–233]. Independent studies also report increased tau phosphorylation in neurons exposed to NO [234], suggesting that NO produced via iNOS-mediated arginine metabolism contributes to AD progression by driving inflammation and potentially influencing tau phosphorylation in neurons. Conversely, M2 microglia express arginase-1, which catalyzes the conversion of arginine into polyamine precursors, promoting anti-inflammatory processes [228]. In the hippocampal region of APP-PS1 mice, Arginase-1+ microglia, which contain more A $\beta$  than iNOS+ microglia, were upregulated and involved in A $\beta$  clearance [235].

In summary, amino acid metabolism plays a pivotal role in regulating microglial functional and phenotypic transitions in both in vivo and in vitro contexts. Among key metabolic pathways, tryptophan metabolism through the kynurenine pathway, mediated by IDO, and arginine metabolism via iNOS and arginase-1, serve as crucial links between microglial activation and AD pathology. However, the precise molecular mechanisms that connect these metabolic reprogramming events to neuroinflammation and A $\beta$ /tau pathology remain incompletely understood. Furthermore, while glutamine metabolism has been extensively investigated, the roles of other amino acids, such as tryptophan and arginine, remain relatively underexplored, highlighting the need for further research to fully elucidate their contributions to AD pathogenesis.

### **Molecular mediators of microglial immunometabolism in AD**

#### **TREM2**

TREM2 is a membrane protein predominantly expressed on microglia in the CNS [35, 161, 236], playing critical roles in the wide-ranging microglial functions, including proliferation, differentiation, phagocytosis, metabolism, survival, lipid handling and inflammatory responses [38, 40, 237–240]. Given its essential role in microglial function and immune homeostasis, genetic variants of TREM2 that lead to loss of function are strongly associated with an increased risk of developing AD [241–243].

Recent studies emphasize that TREM2, which is upregulated in DAM [154, 157], facilitates the transition of microglia to the DAM phenotype [157, 244, 245] while modulating metabolism to support microglial activation and A $\beta$  clearance [35, 39, 157, 245–247]. Research using knockout models has demonstrated that TREM2 activates a metabolic program involving glycolysis, the PPP, and the TCA cycle to support phagocytosis [35]. Glucose uptake, the critical energy source for this metabolic pathway, depends on TREM2 and varies with microglial activation status [133]. Microglia exhibit distinct transcriptomic and metabolic profiles depending on their level of TREM2 expression, with these differences becoming more pronounced near amyloid plaques [248]. TREM2 deficiency in microglia from 5XFAD mice leads to a downregulation of glucose transporters and glycolytic enzymes. This energy deficit, induced by TREM2 deficiency, activates AMPK and impairs mTOR signaling, resulting in mitochondrial damage and increased autophagy [35]. These autophagy-related changes align with the multi-omics analyses in patients with amyloid pathology and AD-like brain organoid models [249], suggesting that alterations in autophagy-related pathways, which are central regulators of cellular metabolism, may be key contributors to AD pathogenesis.

Similar to TREM2-deficient microglia, microglia expressing the R47H variant of TREM2 exhibit metabolic impairments, including reduced mitochondrial oxidative phosphorylation and an inability to undergo glycolytic reprogramming. These metabolic deficits compromise energy production, phagocytosis, and adaptive responses to inflammation, impairing their ability to clear A $\beta$ 42 and further emphasizing the critical role of TREM2 in maintaining microglial metabolic fitness [250]. Likewise, alterations in microglial metabolic pathways have been discovered in the brain of AD patients carrying the R47H variant [251]. Piers et al. found that the observed deficits in microglial metabolic regulation and associated functions in TREM2 variants are caused by dysregulation of the PPAR $\gamma$ /p38MAPK signaling pathway [250]. They showed that by activating these pathways, the metabolic deficits of microglia can be improved, leading to the restoration of A $\beta$  phagocytosis.

TREM2 also plays a crucial role in regulating microglial cholesterol metabolism, particularly during chronic phagocytic challenges. In microglia lacking the TREM2, there is a disruption in the regulation of genes involved in lipid metabolism, resulting in an accumulation of cholesterol ester (CE) and impaired transport of cholesterol within the brain [239, 252]. This indicates that TREM2 is a vital component in maintaining optimal lipid homeostasis in microglia. Knockdown of TREM2 results in increased lipid synthesis and decreased cholesterol

clearance and lipid hydrolysis, further impacting microglial phenotypes. The critical role of TREM2 in regulating cholesterol transport and metabolism in microglia, largely identified through knockout studies, was further corroborated by recent research demonstrating an inverse relationship between TREM2 expression and free cholesterol levels [248]. Microglia expressing high levels of TREM2 showed significantly lower levels of free cholesterol and palmitoylcarnitine compared to their low TREM2 expressing microglia [248], suggesting high lipid catabolism and cholesterol efflux in TREM2-high microglia. Overall, elevated TREM2 expression in microglia correlates with enhanced energetic and metabolic capacities, as well as improved cholesterol management. This suggests a pivotal role for TREM2 in optimizing microglial function through metabolic regulation.

### APOE

Given that TREM2 is a key regulator of microglial lipid metabolism, APOE, another major player in lipid handling, also significantly influences microglial immunometabolism. Upregulation of microglial APOE is a significant feature observed in both AD patients [164, 253] and AD mouse brains [157, 253–255]. A comprehensive multi-omics study revealed that APOE4 significantly alters microglial metabolism and immune responses [256]. APOE4 microglia, isolated from APOE4 KI mice, exhibit increased aerobic glycolysis with elevated HIF-1 $\alpha$  expression and a disrupted TCA cycle compared with APOE3 microglia [70]. These microglia show increased APOE expression and upregulation of genes involved in glucose metabolism, lipid processing, and pro-inflammatory cytokines [70]. These findings are partially consistent with another work that reported that APOE4 significantly impairs the metabolic function of human iPSC-derived microglia, inhibiting both glycolysis and OXPHOS, which in turn hinders their phagocytosis, migration, and overall metabolic activity, while simultaneously exacerbating cytokine secretion [257, 258]. Furthermore, APOE4 5XFAD mice exhibited enriched genes associated with the DAM/MGnD signature in the cortex and hippocampus compared to APOE3 5XFAD mice, indicating an induced microglial transcriptomic switch in response to amyloid pathology [70]. In addition to its effects on glucose metabolism, APOE4 also disrupts lipid homeostasis in microglia, leading to lipid droplet accumulation and impaired lipid processing, particularly in the context of amyloid pathology [256]. Interestingly, deletion of APOE4 in microglia restored the MGnD response to chronic neurodegeneration and promoted neuroprotection by alleviating tau and amyloid pathology [256].



In addition to DAM/MGnD microglia, a recent study identified a novel population of microglia, termed terminally inflammatory microglia (TIM) [259]. TIM, characterized by the expression of inflammatory signals and stress markers, increases with age and APOE4 presence, showing reduced cellular energetics (including depleted citric acid cycle, glycolysis, and sugar metabolism pathways), features of ROS detoxification, and altered amino acid metabolism [259]. An analogous population has been identified in the human AD brain. TIM exhibits impaired A $\beta$  clearance and altered cell–cell communication during aducanumab treatment [259], suggesting that it may contribute to AD risk and pathology, particularly in APOE4 carriers and the elderly, making it a potential therapeutic target.

The brain's primary lipid transporter, APOE, has also been demonstrated to exert isoform-specific effects on lipid and cholesterol metabolism. Microglia derived from human induced pluripotent stem cells (iPSCs) expressing APOE4 were found to contain more LDs [73], supporting recent findings that APOE4 in microglia influences lipid metabolism and potentially contributes to AD pathology. In this recent study by Haney et al. snRNA-seq analysis of AD brain tissue revealed a population of LD-enriched microglia characterized by high ACSL1 expression, an enzyme involved in LD formation [260]. This ACSL1-positive LDAM population was most abundant in patients with the APOE4/4 genotype. In iPSC-derived microglia, A $\beta$  upregulated ACSL1 expression, resulting in triglyceride synthesis and LD accumulation in an APOE-dependent manner. Notably, secreted factors from these LD-laden microglia led to tau phosphorylation and neurotoxicity in an APOE-dependent manner. Recently, the impact of APOE isoform-dependent microglial changes on AD pathogenesis has been investigated using a human microglia xenotransplantation model [261]. In this study, ATAC-seq and RNA-seq were used to distinguish the effects of various APOE isoforms on microglia in AD from epigenomic and transcriptomic perspectives. Significant transcriptomic and epigenetic differences were found in microglia depending on the APOE isoform. APOE4 was shown to potentially increase AD risk by upregulating genes related to lipid accumulation and impairing microglial proliferation, migration, and immune responses [261].

#### **TREM2-APOE interaction**

TREM2 and APOE are pivotal in AD pathogenesis due to their interconnected roles in lipid metabolism, microglial activation, and A $\beta$  clearance [262, 263]. While TREM2 facilitates phagocytosis by recognizing lipid ligands [161, 264], APOE transports lipids and cholesterol, interacting with microglial receptors involved in lipoprotein

metabolism [163]. TREM2 and APOE collaboratively regulate microglial responses to A $\beta$  pathology [159, 262, 263], with TREM2 promoting phagocytosis and APOE facilitating lipid handling. This activation drives the MGnD transition by promoting metabolic and inflammatory reprogramming in microglia, a process accompanied by TGF- $\beta$ 1 suppression [159]. Transcriptomic analyses indicate that TREM2-APOE signaling promotes MGnD-like microglia while downregulating homeostatic genes such as P2ry12, Tmem119, and Tgfr1, suggesting that TGF- $\beta$ 1 suppression may be necessary for MGnD activation, yet its complete loss could impair microglial homeostasis and cognitive function [265].

Recent findings highlight that APOE4 disrupts this balance, leading to excessive lipid droplet accumulation in microglia, particularly in APP-PS1:APOE4 KI models [256]. This metabolic dysregulation mirrors that of TREM2-deficient microglia, where disrupted lipid metabolism and TGF- $\beta$ 1 signaling jointly promote a pro-inflammatory state [266]. TREM2 deficiency disrupts cholesterol homeostasis, leading to cholesteryl ester accumulation and lipid droplet formation, which shifts microglia toward a pro-inflammatory phenotype [266]. Notably, blocking APOE4-TGF- $\beta$ 1 signaling restores MGnD-like phenotypes and enhances A $\beta$  clearance in AD models, highlighting the dual role of TGF- $\beta$ 1 in microglial function—maintaining homeostasis under physiological conditions but potentially impeding MGnD activation and A $\beta$  clearance in AD pathology—thereby underscoring its complex interplay with the TREM2-APOE axis.

Given its crucial role in regulating lipid metabolism and immune function, TREM2 acts as a key receptor for APOE [263, 267, 268], integrating metabolic and inflammatory responses to maintain microglial function in AD pathology. This interaction is essential for coordinating microglial responses to A $\beta$  pathology, further emphasizing its central role in AD progression [12, 269]. Indeed, transcriptomic studies in 5XFAD mice have identified TREM2 and APOE as two of the most upregulated genes in microglia actively engaged in amyloid phagocytosis, underscoring their importance in microglial activation and function in AD pathology [255, 270].

Importantly, TREM2 and APOE are critical for the formation of a microglial barrier surrounding plaques, a structural response that limits amyloid spread and associated toxicity. In the absence of either gene, this protective microglial clustering around A $\beta$  plaques is significantly impaired, highlighting their cooperative role in modulating plaque-associated microglial activity [271]. Microglia expressing APOE4 exhibit a reduced ability to respond to A $\beta$  compared to those expressing APOE3. This deficit is further exacerbated in the

absence of TREM2, leading to a significantly diminished capacity for A $\beta$  phagocytosis [272]. In AD, the TREM2-APOE axis plays a crucial role in microglial activation and A $\beta$  clearance, a function that extends to other neurodegenerative diseases where impaired microglial responses contribute to pathology [159]. A recent study has revealed that externalized phosphatidylserine (ePtdSer), generated from nutrient-deprived neurons surrounding A $\beta$  plaques, serves as the driving force behind TREM2-dependent A $\beta$  phagocytosis in microglia [273]. Intriguingly, microglia expressing APOE4 demonstrated reduced TREM2-dependent A $\beta$  phagocytosis via ePtdSer compared to those expressing APOE3 [273].

While the role of TREM2-APOE interactions in A $\beta$  pathology is relatively well-characterized, their contribution to tau pathology remains less understood, reflecting the broader complexity of AD progression. Recent evidence suggests that APOE4 serves as the primary driver of dysfunction in tau-related pathology [274]. Interestingly, the impact of TREM2 deficiency on tau pathology is dependent on APOE isoform, revealing a more nuanced interplay between these two factors in AD. In TREM2-deficient P301S tauopathy mice expressing murine APOE, neuroinflammation is reduced, and brain atrophy is mitigated, suggesting a neuroprotective role of TREM2 deficiency [275]. However, in P301S mice expressing human APOE4, TREM2 deficiency exacerbates tau pathology and neurodegeneration, despite a reduction in TREM2-dependent microgliosis [274]. Notably, human APOE4 promotes abnormal lipid accumulation in microglial lysosomes, independent of TREM2, further reinforcing its central role in driving tau-related pathology. These findings highlight that the interplay between TREM2 and APOE extends beyond ligand binding, actively shaping microglial responses to neurodegeneration. Given the therapeutic interest in TREM2 modulation, isoform-specific investigations into APOE-TREM2 interactions will be crucial for refining treatment strategies. Furthermore, addressing the interplay of these mechanisms in both amyloid and tau pathologies will be essential for developing comprehensive and effective therapeutic approaches.

APOE-TREM2 interactions are modulated by APOE isoform and lipidation status [263, 268, 276, 277], which may in turn affect microglial function in AD pathology. The degree of APOE lipidation differs among isoforms, with APOE2 exhibiting the highest level and APOE4 showing the lowest [278–281]. TREM2 preferentially binds to non-lipidated APOE, and APOE4 generally displays a higher binding affinity than APOE2 [276, 277], possibly due to its lower lipidation level. Differences in APOE lipidation across isoforms influence not only TREM2 interactions but also key microglial functions,

such as A $\beta$  clearance, lipid homeostasis, and inflammatory responses [282].

A recent study by Guo et al. reported that lipidated APOE2 exhibits reduced LDLR binding, which appears to be a protective mechanism in AD by limiting excessive toxic lipid uptake and preventing microglial overactivation [283]. These findings suggest that APOE lipidation may play a key role in modulating interactions with various APOE receptors, which could be closely linked to the progression of AD pathology.

Phospholipid composition also varies in an APOE isoform-specific manner, contributing to differences in lipidation [272, 284] and microglial responses to A $\beta$  [272]. APOE3 lipoproteins, compared to APOE4, enhance microglial migration, promote A $\beta$  uptake, and induce a stronger activation response, with these effects being more pronounced in the absence of TREM2 [272]. These findings suggest that isoform-specific variations in phospholipid composition contribute to differences in APOE lipidation, which may, in turn, influence APOE-TREM2 interactions and microglial function in A $\beta$  pathology [272].

Lipidated APOE plays a crucial role in A $\beta$  metabolism by forming complexes with A $\beta$ , facilitating its rapid clearance by microglia, and reducing A $\beta$  toxicity [278, 285]. However, poorly lipidated APOE4 exhibits reduced binding affinity to A $\beta$ , leading to slower clearance, increased A $\beta$  toxicity, and heightened inflammation [210, 285–287]. Furthermore, the R47H variant of TREM2 not only shows reduced binding affinity for APOE [263, 267, 277] but also disrupts downstream microglial signaling pathways [288, 289], impairing A $\beta$  clearance and exacerbating neuroinflammation. A $\beta$  interacts with both the lipid-binding and receptor-binding domains of APOE, further complicating APOE-TREM2 interactions and their consequences for microglial function [290, 291].

Given these complexities, a comprehensive understanding of the interactions between APOE and its various receptors, including but not limited to TREM2, is crucial for elucidating the mechanisms underlying microglial dysfunction in AD. In particular, research should focus on how APOE isoform differences, lipidation status, and phospholipid composition influence these receptor interactions and downstream signaling pathways. Gaining these insights may facilitate the development of targeted therapeutic strategies that effectively address both A $\beta$  and tau pathologies in AD.

### **Hypoxia-inducible factors**

Metabolic plasticity refers to the ability of cells to adapt their metabolic processes in response to changing environmental conditions or internal needs. The drivers of metabolic reprogramming that have been most

extensively studied are changes in nutrient availability and oxygen levels [292, 293]. Hypoxia occurs when cells, especially during inflammation, experience increased oxygen consumption [294, 295]. It is a well-established driver of glycolysis, as oxygen deficiency restricts OXPHOS efficiency. Hypoxia-inducible factor 1 (HIF-1) is a heterodimeric transcription factor consisting of HIF-1 $\alpha$  and HIF-1 $\beta$  that plays a crucial role in cellular response to low oxygen conditions, or hypoxia [295, 296]. Under hypoxic conditions, HIF-1 facilitates sustained ATP production by upregulating hexokinase and phosphofructokinase to enhance anaerobic glycolysis and by increasing GLUT1 expression to enhance glucose uptake [295, 297, 298]. HIF-1 $\alpha$  stabilization is not limited to hypoxia but can also be stabilized under normoxic conditions via pro-inflammatory cytokines (e.g., TNF- $\alpha$  and IL-1 $\beta$ ) [299], metabolic intermediates of the TCA cycle (e.g., succinate and fumarate) [300], and NO [301, 302]. These factors inhibit prolyl hydroxylase (PHD) activity, preventing HIF-1 $\alpha$  degradation [300, 301]. Such normoxic regulatory mechanisms, observed in macrophages [297, 303], may also influence microglial metabolism.

Hypoxia and inflammation are closely linked, with HIF-1 serving as a key switch in glycolytic reprogramming and acting as a crucial transcriptional regulator of immunity and inflammation [294, 295, 298, 304]. Baik et al. demonstrated that primary microglia exposed to A $\beta$  induce metabolic reprogramming from OXPHOS to glycolysis via the mTOR-HIF-1 $\alpha$  pathway, leading to an acute inflammatory state [46]. However, chronic A $\beta$  exposure results in metabolic deficits and downregulation of the mTOR-HIF-1 $\alpha$  pathway, impairing microglial function and reducing their phagocytic capacity [46]. Restoration of this pathway via IFN- $\gamma$  treatment in 5xFAD mice reactivated microglial immune responses and enhanced phagocytic function, suggesting metabolic modulation as a potential therapeutic approach for AD [46]. Conversely, Yang et al. proposed a contrasting strategy that targets the same pathway but employs an opposing approach [305]. They developed an immunometabolic reprogramming nanomodulator (GAF NPs) that inhibits the mTOR-HIF-1 $\alpha$  pathway, shifting microglial metabolism from glycolysis to OXPHOS and promoting an M2 anti-inflammatory phenotype. This inhibition was accompanied by GLUT1 suppression, reducing glucose uptake and further limiting glycolytic activity. By dampening excessive inflammatory responses, this strategy led to cognitive improvement and decreased A $\beta$  accumulation in APP-PS1 mice [305]. A recent study introduced a peptide-drug conjugates (PDCs)-based nanoplateform designed to enhance microglial function by restoring the mTOR-HIF-1 $\alpha$  pathway, particularly in later stages of AD pathology [306]. Notably, this study used the same

APP-PS1 model as Yang et al., yet adopted a strategy similar to Baik et al. by upregulating HIF-1 $\alpha$ . The researchers found that the activation state of the mTOR-HIF-1 $\alpha$  pathway was highly dependent on disease progression. In APP-PS1 mice, HIF-1 $\alpha$  and p-mTOR were significantly elevated at 7 months of age, aligning with the timing of Yang et al.'s intervention, which aimed to suppress this pathway. However, by 9 months of age, a sharp decline in HIF-1 $\alpha$  and mTOR activity was observed, indicative of an immune-suppressed state resembling the chronic phase described by Baik et al. In this context, PDCs treatment successfully restored mTOR-HIF-1 $\alpha$  signaling, leading to improved microglial function and neuroprotection in advanced-stage AD [306]. These findings reinforce the notion that disease progression and microglial metabolic states can vary depending on the animal model and experimental time points. The opposing strategies proposed by Baik et al. and Yang et al. may not be mutually exclusive but rather context-dependent, influenced by the stage of AD pathology at which the intervention is applied. Therefore, the therapeutic success of targeting the HIF-1 $\alpha$ -mTOR pathway may depend on precise timing to maximize its efficacy, underscoring the importance of understanding microglial metabolic transitions across different disease stages for the development of effective immunometabolic interventions in AD.

In 5XFAD mice, microglia exhibit distinct transcriptional profiles depending on whether they contain A $\beta$  [307]. In 5XFAD mice, microglia containing A $\beta$  exhibited a distinct transcriptional profile characterized by an active HIF-1 $\alpha$  regulon. Notably, microglia with this transcriptional signature are also increased in postmortem AD brains [307]. This upregulation of HIF-1 $\alpha$  may support the enhanced metabolic and phagocytic activities required to manage the pathological environment induced by A $\beta$  plaques in AD. Similar findings have been observed in A $\beta$ -associated microglia of APP-PS1 mice, where activation of the HIF-1 pathway and transcription of mitochondrial-related genes were noted. This was accompanied by mitochondrial elongation, an adaptive response to support aerobic respiration under low nutrient and oxygen conditions [308]. However, sustained HIF-1 activation or prolonged hypoxia can induce microglial quiescence, diminishing their responsiveness to A $\beta$ . This low-reactivity state diminishes microglial clustering around A $\beta$  plaques, ultimately exacerbating A $\beta$  pathology [308].

Additionally, the complement C3a receptor (C3aR) signaling pathway has been implicated in microglial metabolic regulation, in part through its influence on HIF-1 $\alpha$  activity [309]. Under hypoxia-mimicking conditions, C3aR-deficient microglia exhibited reduced HIF-1 $\alpha$  levels, which was accompanied by the downregulation of

its glycolysis-promoting target genes, including *PDK1*, *PFKL*, and *PFK2* [309]. Notably, these microglia exhibited resistance to hypoxia mimetic-induced metabolic changes and lipid droplet accumulation. Consistent with these findings, C3aR expression is elevated around A $\beta$  plaques in APP-KI mice, where microglia with high C3aR expression show metabolic impairments characterized by increased HIF-1 $\alpha$  signaling and disrupted lipid metabolism, ultimately affecting their response to A $\beta$  pathology [309]. However, C3aR deficiency in these mice restored lipid profiles, reduced lipid droplet accumulation, and enhanced microglial phagocytic capacity. These changes were accompanied by a decrease in A $\beta$  burden and a modest improvement in cognitive function [309]. This suggests that the C3aR/HIF-1 $\alpha$  axis plays a critical role in regulating microglial metabolic adaptation and may be a potential therapeutic target in AD.

Further evidence linking HIF-1 $\alpha$  to microglial metabolism comes from studies implicating key AD risk genes, such as *TREM2* and *APOE4*, in its regulation. In an AD mouse model, *TREM2* deficiency leads to significant metabolic alterations in microglia, including reduced mitochondrial mass and downregulated HIF-1 $\alpha$  expression [35]. This suggests that *TREM2* plays a vital role in maintaining microglial metabolic fitness through regulation of HIF-1 $\alpha$ -mediated glycolytic pathways. Conversely, *APOE4* promotes a glycolytic phenotype in microglia via increased HIF-1 $\alpha$  activation, particularly in aging. Aged *APOE4* microglia exhibit elevated aerobic glycolysis, accumulation of glycolytic metabolites, and the emergence of a DAM-like microglial subpopulation [70]. This HIF-1 $\alpha$ -high, DAM-like phenotype is linked to disrupted cholesterol metabolism, further highlighting the interplay between *APOE4*, microglial metabolic states, and AD pathology [70].

These findings underscore the complex and context-dependent role of HIF-1 $\alpha$  in AD pathology. The effects of HIF-1 $\alpha$  modulation likely depend on microglial activation states, disease progression, and cellular metabolism, necessitating a nuanced approach when considering it as a therapeutic target.

### AMPK

Adenosine monophosphate-activated protein kinase (AMPK), a serine/threonine protein kinase, serves as an energy sensor and master regulator, playing a pivotal role in maintaining cellular energy homeostasis [310]. Beyond its role as an energy sensor, AMPK coordinates multiple metabolic pathways, including glucose metabolism, lipid metabolism, and protein synthesis, to adapt cellular function under energy stress conditions [310–312]. Classical activation of AMPK occurs when the ratio of adenine nucleotides (AMP to ATP and/or ADP to ATP) within

the cell increases. Upon activation, AMPK restores ATP levels during metabolic stress by simultaneously inhibiting ATP-consuming biosynthetic pathways and activating ATP-generating catabolic pathways [78]. Furthermore, AMPK phosphorylates multiple transcription factors such as FOXO3, CREB, and p300 that play crucial roles in biosynthetic pathways and metabolism regulation [310]. Through these mechanisms, AMPK rapidly restores energy balance and transcriptionally modulates cell metabolism in response to long-term energy deficits [313].

Autophagy plays a crucial role in maintaining microglial homeostasis by enabling cells to adapt to metabolic stress through the degradation and recycling of cellular components [314]. AMPK activation promotes autophagy by inhibiting mTOR, a key regulator of cell growth that suppresses autophagy in nutrient-rich conditions [315]. This AMPK-mTOR balance is particularly crucial in microglial function, as it regulates inflammatory responses and microglial polarization [316–318]. In neurodegenerative diseases such as AD, persistent metabolic stress and inflammation impair autophagic flux, leading to the accumulation of dysfunctional proteins and organelles [314, 319]. By modulating catabolic and anabolic processes [320], AMPK signaling contributes to maintaining cellular adaptation and homeostasis, potentially influencing A $\beta$  clearance and neuroinflammation dynamics.

As AMPK activation exerts anti-inflammatory effects [321], metabolic changes in response to immune stimuli may suppress AMPK function, leading to immune responses such as pro-inflammatory cytokine secretion and inflammasome activation. As late-stage AD progresses, chronic low-level inflammation persists, exacerbating neurotoxicity. A $\beta$ -driven neuroinflammation is partly mediated through the NLRP3 inflammasome [322], which is activated in microglia following AMPK inhibition and Syk signaling activation. Research shows that AMPK deactivation leads to metabolic dysregulation, mitochondrial fragmentation, and reactive oxygen species production, thereby promoting NLRP3 inflammasome activation [323]. As a central regulator of cellular metabolism, AMPK influences multiple metabolic pathways, including TCA cycle, FAO, cholesterol homeostasis, and glycolysis, to maintain energy balance. As these metabolic processes are also involved in NLRP3 inflammasome activation [206, 324–326], AMPK dysfunction may further exacerbate neuroinflammatory responses in AD by disrupting metabolic homeostasis. In this context, NSAIDs or metformin, which have a direct anti-inflammatory function and enhance AMPK activity, may serve as potential adjuvant treatments to modulate microglial immunometabolism [323, 327–330].



Interestingly, INPP5D, a microglial gene identified through GWAS as a late-onset AD risk factor [13, 67, 331], has been implicated in autophagy and NLRP3 inflammasome activation [332], both of which are closely linked to AMPK signaling and metabolic regulation [333, 334]. While INPP5D expression is elevated in amyloid plaque-associated microglia [332, 335], its functional role appears to be context-dependent, with potential contributions to both protective and pathological processes [336–338]. A recent study reported that the functional decline caused by INPP5D reduction is associated with disrupted autophagy flux and increased activation of the NLRP3 inflammasome [332]. However, INPP5D can modulate the downstream activity of multiple receptors involved in microglial activation and function [337], and AMPK signaling can influence intracellular pathways that are also affected by INPP5D, necessitating further research to clarify its role in AD pathogenesis.

AMPK plays a central role in regulating cellular metabolism, influencing multiple pathways including glucose metabolism, lipid homeostasis, mitochondrial function, energy production and utilization, and protein synthesis [311]. Through these metabolic networks, AMPK modulates cellular stress responses, coordinating autophagy, mitophagy, and lysosomal function while also shaping inflammatory pathways such as NLRP3 inflammasome activation [318]. Given its ability to integrate metabolic and immune signaling, AMPK represents a key node linking energy homeostasis to neuroinflammatory regulation in AD pathogenesis.

These multifaceted roles of AMPK in regulating microglial inflammatory responses offer insights into its function and impact on AD pathogenesis. As a central metabolic regulator, AMPK influences various processes, including autophagy, mitochondrial homeostasis, and immune responses, making it a compelling target for therapeutic intervention [339–342]. However, AMPK operates within a complex signaling network involving LOAD risk genes such as TREM2 and other metabolic regulators [343, 344], meaning its modulation could have varying effects depending on cellular context, disease stage, and microglial activation states [57, 341]. Directly targeting AMPK may lead to unintended consequences, highlighting the need to carefully consider whether modulating upstream regulators or downstream pathways—such as autophagy and NLRP3 inflammasome activation—offers a more viable therapeutic strategy for AD.

### Hexokinase

Hexokinases (HKs) are the initial rate-limiting enzymes in glucose metabolism, catalyzing the phosphorylation of glucose to glucose-6-phosphate (G-6-P), thereby

initiating the first committed step in glycolysis. Recent studies on tissue-resident macrophages, including microglia, have highlighted the critical role of hexokinase in glycolysis and its influence on mitochondrial function, which is dynamically regulated by inflammatory activation and plays a key role in immune responses [345–347]. Mammals express five isoforms of hexokinase (HK), with the four primary isoforms being HK1, HK2, HK3, and HK4 (also known as glucokinase), along with an additional isoform, HKDC1. Among these, HK1, HK2, and HK4 are localized to the outer mitochondrial membrane (OMM), where they interact with voltage-dependent anion channels (VDACs) to facilitate ATP-dependent glucose phosphorylation [348]. In the brain, HK1, HK2, and HK3 are expressed, but notably, HK2 levels were significantly elevated in AD patients [50, 75, 349]. RNA-seq analyses across various brain cell types, supported by *in situ* hybridization and immunoblotting, revealed that HK2 is selectively enriched in microglia, both in murine and human brains, compared to other brain cells. Moreover, HK2 expression was markedly elevated in microglia undergoing immune activation or associated with disease states [349]. Surprisingly, contrary to the expectation that depleting HK2—the initial enzyme in energy metabolism—would suppress metabolism, its deletion instead increased microglial energy demand and enhanced phagocytic activity [75, 349]. These findings suggest that HK2 overexpression in microglia does not efficiently support the energy requirements for phagocytosis. Interestingly, under chronic conditions such as AD, genetic or pharmacological depletion of HK2 induced a metabolic shift in microglia from glucose-dependent metabolism to lipid utilization. This metabolic adaptation enhanced ATP production, thereby delaying the onset of AD symptoms in mouse models [75, 350]. Beyond its role in glycolysis, HK2 also has non-metabolic functions [351–353], contributing to microglial inflammatory responses. Its impact on disease progression appears to be gene-dosage-dependent, as complete deletion of HK2 exacerbates inflammation and mitochondrial dysfunction, whereas partial reduction (haploinsufficiency) alleviates AD pathogenesis in AD model mice [50].

These findings suggest that the balance between HK2's metabolic and non-metabolic functions is tightly regulated in response to physiological and pathological stimuli. Further research into the multifaceted roles of HK2 will be essential to understanding the reciprocal regulation of metabolic and inflammatory processes in microglia and their contributions to AD pathogenesis and progression.

## Conclusion and future directions

Recent advances in multi-omics technologies, including single-cell genomics, transcriptomics, proteomics, metabolomics, and epigenomics, have unveiled the remarkable heterogeneity of microglial states under both physiological and pathological conditions. This diversity reflects the dynamic nature of microglia, which continuously adapt to genetic and environmental influences throughout aging and neurodegenerative processes (Fig. 1, Table 1). While this complexity poses challenges for drug development, it also presents new opportunities for therapeutic intervention in Alzheimer's disease (AD). Given the critical role of metabolic regulation in microglial function, targeting metabolic pathways has emerged as a promising therapeutic approach for AD [43, 45, 92, 305]. However, the intricate interplay between microglial transcriptional states, metabolic adaptations, and functional roles necessitates a more refined therapeutic strategy. The identification of distinct microglial subpopulations, each exhibiting unique metabolic profiles and functional states [354–356], highlights the need for a comprehensive and context-dependent understanding of microglial behavior in AD. Despite these insights, methodological limitations—such as incomplete capture of microglial states and insufficient spatial resolution in pathology assessment—continue to impede progress.

A recent study has provided a detailed gene expression map across multiple brain regions in both AD patients and healthy individuals [357]. This study identified cell type- and region-specific genetic alterations in AD, revealing transcriptomic differences associated with diverse pathological factors [357]. These findings underscore the complexity of microglial metabolic networks and emphasize the necessity of considering genetic predispositions, environmental influences, and comorbidities in the development of targeted therapeutic strategies.

Growing evidence suggests a bidirectional relationship between AD pathology and microglial metabolic dysfunction. A $\beta$  and tau aggregates drive metabolic reprogramming, impairing microglial immune function [46, 53, 86, 87, 358]. Conversely, pre-existing metabolic disturbances may predispose microglia to a dysfunctional state, further exacerbating AD progression [60, 359, 360]. This interplay may differ between sporadic and familial AD, where familial AD is primarily driven by early A $\beta$  accumulation, whereas sporadic AD appears to be more influenced by systemic metabolic dysfunction and neuroinflammation [361–363]. Moreover, metabolic disorders such as obesity and type 2 diabetes, which are closely linked to an increased risk of AD [364–369], along with their associated metabolic hormones, may influence brain immunity and microglial function [370–374]. Individuals with these metabolic conditions may exhibit

reduced microglial resilience to pathological stimuli such as A $\beta$ , thereby impairing their ability to mount an effective response. This diminished adaptive capacity may lead to defective immune responses and compromised A $\beta$  clearance, ultimately accelerating AD progression.

Moving forward, research efforts should aim to clarify the mechanistic links between metabolic pathways and microglial dysfunction, as well as to identify novel therapeutic targets that restore microglial homeostasis. A deeper understanding of how systemic metabolic status influences microglial function will be essential for designing effective AD treatments. Additionally, integrating these findings within the spatial and temporal framework of disease progression is crucial for the development of precision-targeted therapies. Addressing these challenges may pave the way for metabolic-based therapeutic interventions, offering a transformative approach to AD treatment.

## Abbreviations

A $\beta$	Amyloid-beta
AD	Alzheimer's disease
AMPK	Adenosine monophosphate-activated protein kinase
APOE	Apolipoprotein E
APOE4	APOE- $\epsilon$ 4
CE	Cholesterol ester
DAMs	Disease-associated microglia
FAO	Fatty acid oxidation
FAs	Fatty acids
FFAs	Free fatty acids
G-6-P	Glucose-6-phosphate
GAC	Glutaminase C
GLS	Glutaminase
GLUTs	Glucose transporters
HIF-1	Hypoxia-inducible factor 1
HK2	Hexokinase II
iMGL	iPSC-derived microglia
iPSCs	Induced pluripotent stem cells
LD	Lipid droplet
LDAM	Lipid-droplet-accumulating microglia
LDs	Lipid droplets
LPL	Lipoprotein Lipase
MnD	Microglial neurodegenerative phenotype
OXPHOS	Oxidative phosphorylation
PPP	Pentose phosphate pathway
ROS	Reactive oxygen species
scRNA-seq	Single-cell RNA sequencing
TAGs	Triacylglycerols
TCA	Tricarboxylic acid
TIM	Termed late-stage inflammatory microglia
TREM2	Triggering receptor expressed on myeloid cells 2

## Acknowledgements

A figure of the article was created with BioRender.com.

## Authors' contributions

E.S.J, H.C and I. M.-J wrote the manuscript. All authors read and approved the final manuscript.

## Funding

This work was supported by a grant from the Korea Health Technology R&D Project through the Korea Health Industry Development Institute (KHIDI) and Korea Dementia Research Center (KDRC), funded by the Ministry of Health & Welfare and Ministry of Science and ICT, Republic of Korea (grant numbers: RS-2020-KH106747 and RS-2020-KH106773). The work was also supported



by grant from the National Research Foundation of Korea (grant number: RS-2023-00274420).

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

The authors declare that they have no competing interests.

Received: 1 September 2024 Accepted: 5 March 2025

Published online: 27 March 2025

## References

- Ferrari C, Sorbi S. The complexity of Alzheimer's disease: an evolving puzzle. *Physiol Rev*. 2021;101:1047–81.
- Jorfi M, Maaser-Hecker A, Tanzi RE. The neuroimmune axis of Alzheimer's disease. *Genome Med*. 2023;15:6.
- Duara R, Barker W. Heterogeneity in Alzheimer's disease diagnosis and progression rates: implications for therapeutic trials. *Neurotherapeutics*. 2022;19:8–25.
- Jellinger KA. Recent update on the heterogeneity of the Alzheimer's disease spectrum. *J Neural Transm (Vienna)*. 2022;129:1–24.
- Al-Kuraishy HM, Jabir MS, Albuhadily AK, Al-Gareeb AI, Rafeeq MF. The link between metabolic syndrome and Alzheimer disease: a mutual relationship and long rigorous investigation. *Ageing Res Rev*. 2023;91:102084.
- Ezkurdia A, Ramirez MJ, Solas M. Metabolic syndrome as a risk factor for Alzheimer's disease: a focus on insulin resistance. *Int J Mol Sci*. 2023;24:4354.
- Zuin M, Roncon L, Passaro A, Cervellati C, Zuliani G. Metabolic syndrome and the risk of late onset Alzheimer's disease: an updated review and meta-analysis. *Nutr Metab Cardiovasc Dis*. 2021;31:2244–52.
- Neeland IJ, Lim S, Tchernof A, Gastaldelli A, Rangaswami J, Ndumele CE, Powell-Wiley TM, Despres JP. Metabolic syndrome. *Nat Rev Dis Primers*. 2024;10:77.
- Kabir MT, Uddin MS, Mamun AA, Jeandet P, Aleya L, Mansouri RA, Ashraf GM, Mathew B, Bin-Jumah MN, Abdel-Daim MM. Combination drug therapy for the management of Alzheimer's disease. *Int J Mol Sci*. 2020;21:3272.
- Cummings JL, Osse AML, Kinney JW, Cammann D, Chen J. Alzheimer's disease: combination therapies and clinical trials for combination therapy development. *CNS Drugs*. 2024;38:613–24.
- Golde TE. Disease-modifying therapies for Alzheimer's disease: more questions than answers. *Neurotherapeutics*. 2022;19:209–27.
- Efthymiou AG, Goate AM. Late onset Alzheimer's disease genetics implicates microglial pathways in disease risk. *Mol Neurodegener*. 2017;12:43.
- Wightman DP, Jansen IE, Savage JE, Shadrin AA, Bahrami S, Holland D, Rongve A, Borte S, Winsvold BS, Drange OK, et al. A genome-wide association study with 1,126,563 individuals identifies new risk loci for Alzheimer's disease. *Nat Genet*. 2021;53:1276–82.
- Sudwatts A, Thinakaran G. Alzheimer's genes in microglia: a risk worth investigating. *Mol Neurodegener*. 2023;18:90.
- Podlesny-Drabiniok A, Marcora E, Goate AM. Microglial phagocytosis: a disease-associated process emerging from Alzheimer's disease genetics. *Trends Neurosci*. 2020;43:965–79.
- Villegas-Llerena C, Phillips A, Garcia-Reitboeck P, Hardy J, Pocock JM. Microglial genes regulating neuroinflammation in the progression of Alzheimer's disease. *Curr Opin Neurobiol*. 2016;36:74–81.
- Dubbelaar ML, Kracht L, Eggen BJL, Boddeke E. The kaleidoscope of microglial phenotypes. *Front Immunol*. 2018;9:1753.
- Li Q, Barres BA. Microglia and macrophages in brain homeostasis and disease. *Nat Rev Immunol*. 2018;18:225–42.
- Yin J, Valin KL, Dixon ML, Leavenworth JW. The role of microglia and macrophages in CNS homeostasis, autoimmunity, and cancer. *J Immunol Res*. 2017;2017:5150678.
- Ransohoff RM, Brown MA. Innate immunity in the central nervous system. *J Clin Invest*. 2012;122:1164–71.
- Ma M, Jiang W, Zhou R. DAMPs and DAMP-sensing receptors in inflammation and diseases. *Immunity*. 2024;57:752–71.
- Shabab T, Khanabdali R, Moghadamtousi SZ, Kadir HA, Mohan G. Neuroinflammation pathways: a general review. *Int J Neurosci*. 2017;127:624–33.
- Liu P, Wang Y, Sun Y, Peng G. Neuroinflammation as a potential therapeutic target in Alzheimer's disease. *Clin Interv Aging*. 2022;17:665–74.
- Zhang W, Xiao D, Mao Q, Xia H. Role of neuroinflammation in neurodegeneration development. *Signal Transduct Target Ther*. 2023;8:267.
- Fu WY, Wang X, Ip NY. Targeting neuroinflammation as a therapeutic strategy for Alzheimer's disease: mechanisms, drug candidates, and new opportunities. *ACS Chem Neurosci*. 2019;10:872–9.
- Wong-Guerra M, Calfio C, Maccioni RB, Rojo LE. Revisiting the neuroinflammation hypothesis in Alzheimer's disease: a focus on the drugability of current targets. *Front Pharmacol*. 2023;14:1161850.
- Shemer A, Erny D, Jung S, Prinz M. Microglia plasticity during health and disease: an immunological perspective. *Trends Immunol*. 2015;36:614–24.
- Augusto-Oliveira M, Arrifano GP, Delage CI, Tremblay ME, Crespo-Lopez ME, Verkhratsky A. Plasticity of microglia. *Biol Rev Camb Philos Soc*. 2022;97:217–50.
- Yang S, Qin C, Hu ZW, Zhou LQ, Yu HH, Chen M, Bosco DB, Wang W, Wu LJ, Tian DS. Microglia reprogram metabolic profiles for phenotype and function changes in central nervous system. *Neurobiol Dis*. 2021;152:105290.
- Bernier LP, York EM, Kamyabi A, Choi HB, Weillinger NL, MacVicar BA. Microglial metabolic flexibility supports immune surveillance of the brain parenchyma. *Nat Commun*. 2020;11:1559.
- Aldana BI. Microglia-specific metabolic changes in neurodegeneration. *J Mol Biol*. 2019;431:1830–42.
- O'Neill LA, Pearce EJ. Immunometabolism governs dendritic cell and macrophage function. *J Exp Med*. 2016;213:15–23.
- Ganeshan K, Chawla A. Metabolic regulation of immune responses. *Annu Rev Immunol*. 2014;32:609–34.
- Li RY, Qin Q, Yang HC, Wang YY, Mi YX, Yin YS, Wang M, Yu CJ, Tang Y. TREM2 in the pathogenesis of AD: a lipid metabolism regulator and potential metabolic therapeutic target. *Mol Neurodegener*. 2022;17:40.
- Ulland TK, Song WM, Huang SC, Ulrich JD, Sergushichev A, Beatty WL, Loboda AA, Zhou Y, Cairns NJ, Kambal A, et al. TREM2 maintains microglial metabolic fitness in Alzheimer's disease. *Cell*. 2017;170(649–663):e613.
- Brandon JA, Farmer BC, Williams HC, Johnson LA. APOE and Alzheimer's disease: neuroimaging of metabolic and cerebrovascular dysfunction. *Front Aging Neurosci*. 2018;10:180.
- Liao F, Yoon H, Kim J. Apolipoprotein E metabolism and functions in brain and its role in Alzheimer's disease. *Curr Opin Lipidol*. 2017;28:60–7.
- Ulrich JD, Ulland TK, Colonna M, Holtzman DM. Elucidating the role of TREM2 in Alzheimer's disease. *Neuron*. 2017;94:237–48.
- Zheng H, Cheng B, Li Y, Li X, Chen X, Zhang YW. TREM2 in Alzheimer's Disease: microglial survival and energy metabolism. *Front Aging Neurosci*. 2018;10:395.
- Painter MM, Atagi Y, Liu CC, Rademakers R, Xu H, Fryer JD, Bu G. TREM2 in CNS homeostasis and neurodegenerative disease. *Mol Neurodegener*. 2015;10:43.
- Faraji P, Kuhn H, Ahmadian S. Multiple roles of apolipoprotein E4 in oxidative lipid metabolism and ferroptosis during the pathogenesis of Alzheimer's disease. *J Mol Neurosci*. 2024;74:62.
- Huang Y, Mahley RW. Apolipoprotein E: structure and function in lipid metabolism, neurobiology, and Alzheimer's diseases. *Neurobiol Dis*. 2014;72(Pt A):3–12.
- Benarroch E. What is the role of microglial metabolism in inflammation and neurodegeneration? *Neurology*. 2022;99:99–105.
- Borst K, Schwabenland M, Prinz M. Microglia metabolism in health and disease. *Neurochem Int*. 2019;130:104331.

45. Fumagalli M, Lombardi M, Gressens P, Verderio C. How to reprogram microglia toward beneficial functions. *Glia*. 2018;66:2531–49.
46. Baik SH, Kang S, Lee W, Choi H, Chung S, Kim JI, Mook-Jung I. A breakdown in metabolic reprogramming causes microglia dysfunction in Alzheimer's disease. *Cell Metab*. 2019;30(493–507):e496.
47. Olesova D, Dobesova D, Majerova P, Brumarova R, Kvasnicka A, Kouril S, Stevens E, Hanes J, Fialova L, Michalicova A, et al. Changes in lipid metabolism track with the progression of neurofibrillary pathology in tauopathies. *J Neuroinflammation*. 2024;21:78.
48. Xia D, Lianoglou S, Sandmann T, Calvert M, Suh JH, Thomsen E, Dugas J, Pizzo ME, DeVos SL, Earr TK, et al. Novel App knock-in mouse model shows key features of amyloid pathology and reveals profound metabolic dysregulation of microglia. *Mol Neurodegener*. 2022;17:41.
49. Shi Q, Chang C, Saliba A, Bhat MA. Microglial mTOR activation upregulates Trem2 and enhances beta-amyloid plaque clearance in the 5XFAD Alzheimer's disease model. *J Neurosci*. 2022;42:5294–313.
50. Codocedo JF, Mera-Reina C, Bor-Chian Lin P, Fallen PB, Puntambekar SS, Casali BT, Jury-Garfe N, Martinez P, Lasagna-Reeves CA, Landreth GE. Therapeutic targeting of immunometabolism reveals a critical reliance on hexokinase 2 dosage for microglial activation and Alzheimer's progression. *Cell Rep*. 2024;43:114488.
51. Sadeghdoust M, Das A, Kaushik DK. Fueling neurodegeneration: metabolic insights into microglia functions. *J Neuroinflammation*. 2024;21:300.
52. Chen H, Guo Z, Sun Y, Dai X. The immunometabolic reprogramming of microglia in Alzheimer's disease. *Neurochem Int*. 2023;171:105614.
53. Zhao Y, Liu B, Wang J, Xu L, Yu S, Fu J, Yan X, Su J. Abeta and tau regulate microglia metabolism via exosomes in Alzheimer's disease. *Biomedicine*. 1800;2022:10.
54. Rhein V, Song X, Wiesner A, Ittner LM, Baysang G, Meier F, Ozmen L, Bluethmann H, Drose S, Brandt U, et al. Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proc Natl Acad Sci U S A*. 2009;106:20057–62.
55. Edison P. Neuroinflammation, microglial activation, and glucose metabolism in neurodegenerative diseases. *Int Rev Neurobiol*. 2020;154:325–44.
56. Zhang S, Lachance BB, Mattson MP, Jia X. Glucose metabolic crosstalk and regulation in brain function and diseases. *Prog Neurobiol*. 2021;204:102089.
57. Cheng J, Zhang R, Xu Z, Ke Y, Sun R, Yang H, Zhang X, Zhen X, Zheng LT. Early glycolytic reprogramming controls microglial inflammatory activation. *J Neuroinflammation*. 2021;18:129.
58. Bernier LP, York EM, MacVicar BA. Immunometabolism in the brain: how metabolism shapes microglial function. *Trends Neurosci*. 2020;43:854–69.
59. Ghosh S, Castillo E, Frias ES, Swanson RA. Bioenergetic regulation of microglia. *Glia*. 2018;66:1200–12.
60. Mosher KI, Wyss-Coray T. Microglial dysfunction in brain aging and Alzheimer's disease. *Biochem Pharmacol*. 2014;88:594–604.
61. Yamamoto S, Masuda T. Lipid in microglial biology - from material to mediator. *Inflamm Regen*. 2023;43:38.
62. Loving BA, Bruce KD. Lipid and lipoprotein metabolism in microglia. *Front Physiol*. 2020;11:393.
63. Chausse B, Kakimoto PA, Kann O. Microglia and lipids: how metabolism controls brain innate immunity. *Semin Cell Dev Biol*. 2021;112:137–44.
64. Leyrolle Q, Laye S, Nadjar A. Direct and indirect effects of lipids on microglia function. *Neurosci Lett*. 2019;708:134348.
65. Marschallinger J, Iram T, Zardeneta M, Lee SE, Lehallier B, Haney MS, Pluvinage JV, Mathur V, Hahn O, Morgens DW, et al. Lipid-droplet-accumulating microglia represent a dysfunctional and proinflammatory state in the aging brain. *Nat Neurosci*. 2020;23:194–208.
66. Prakash P, Manchanda P, Paouri E, Bisht K, Sharma K, Rajpoot J, Wendt V, Hossain A, Wijewardhane PR, Randolph CE, et al. Amyloid beta Induces Lipid Droplet-Mediated Microglial Dysfunction in Alzheimer's Disease. *bioRxiv* 2024.
67. Kunkle BW, Grenier-Boley B, Sims R, Bis JC, Damotte V, Naj AC, Boland A, Vronskaya M, van der Lee SJ, Amlie-Wolf A, et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates Abeta, tau, immunity and lipid processing. *Nat Genet*. 2019;51:414–30.
68. Kelly B, Pearce EL. Amino acids: how amino acids support immunity. *Cell Metab*. 2020;32:154–75.
69. Hollinger KR, Zhu X, Khoury ES, Thomas AG, Liaw K, Tallon C, Wu Y, Prchalova E, Kamiya A, Rojas C, et al. Glutamine antagonist JHU-083 normalizes aberrant hippocampal glutaminase activity and improves cognition in APOE4 Mice. *J Alzheimers Dis*. 2020;77:437–47.
70. Lee S, Devanney NA, Golden LR, Smith CT, Schwartz JL, Walsh AE, Clarke HA, Goulding DS, Allenger EJ, Morillo-Segovia G, et al. APOE modulates microglial immunometabolism in response to age, amyloid pathology, and inflammatory challenge. *Cell Rep*. 2023;42:112196.
71. Cheng-Hathaway PJ, Reed-Geaghan EG, Jay TR, Casali BT, Bemiller SM, Puntambekar SS, von Saucken VE, Williams RY, Karlo JC, Moutinho M, et al. The Trem2 R47H variant confers loss-of-function-like phenotypes in Alzheimer's disease. *Mol Neurodegener*. 2018;13:29.
72. Wang N, Wang M, Jeevaratnam S, Rosenberg C, Ikezu TC, Shue F, Doss SV, Alnobani A, Martens YA, Wren M, et al. Opposing effects of apoE2 and apoE4 on microglial activation and lipid metabolism in response to demyelination. *Mol Neurodegener*. 2022;17:75.
73. Victor MB, Leary N, Luna X, Meharena HS, Scannell AN, Bozzelli PL, Samaan G, Murdock MH, von Maydell D, Effenberger AH, et al. Lipid accumulation induced by APOE4 impairs microglial surveillance of neuronal-network activity. *Cell Stem Cell*. 2022;29(1197–1212):e1198.
74. Eskandari-Sedighi G, Blurton-Jones M. Microglial APOE4: more is less and less is more. *Mol Neurodegener*. 2023;18:99.
75. Leng L, Yuan Z, Pan R, Su X, Wang H, Xue J, Zhuang K, Gao J, Chen Z, Lin H, et al. Microglial hexokinase 2 deficiency increases ATP generation through lipid metabolism leading to beta-amyloid clearance. *Nat Metab*. 2022;4:1287–305.
76. Fang J, Luo S, Lu Z. HK2: Gatekeeping microglial activity by tuning glucose metabolism and mitochondrial functions. *Mol Cell*. 2023;83:829–31.
77. Semenza GL. HIF-1 mediates metabolic responses to intratumoral hypoxia and oncogenic mutations. *J Clin Invest*. 2013;123:3664–71.
78. Hardie DG, Ross FA, Hawley SA. AMPK: a nutrient and energy sensor that maintains energy homeostasis. *Nat Rev Mol Cell Biol*. 2012;13:251–62.
79. Roberts DJ, Miyamoto S. Hexokinase II integrates energy metabolism and cellular protection: acting on mitochondria and TORC1 to autophagy. *Cell Death Differ*. 2015;22:364.
80. Mergenthaler P, Lindauer U, Dienel GA, Meisel A. Sugar for the brain: the role of glucose in physiological and pathological brain function. *Trends Neurosci*. 2013;36:587–97.
81. Lauro C, Limatola C. Metabolic reprogramming of microglia in the regulation of the innate inflammatory response. *Front Immunol*. 2020;11:493.
82. Bielani JP, Sun D. Significance of microglial energy metabolism in maintaining brain homeostasis. *Transl Stroke Res*. 2023;14:435–7.
83. Kierans SJ, Taylor CT. Glycolysis: a multifaceted metabolic pathway and signaling hub. *J Biol Chem*. 2024;300:107906.
84. Kukuryga MA, Rosset S, Titov DV. The Warburg Effect is the result of faster ATP production by glycolysis than respiration. *Proc Natl Acad Sci U S A*. 2024;121:e2409509121.
85. Pearce EL, Pearce EJ. Metabolic pathways in immune cell activation and quiescence. *Immunity*. 2013;38:633–43.
86. Lu J, Zhou W, Dou F, Wang C, Yu Z. TRPV1 sustains microglial metabolic reprogramming in Alzheimer's disease. *EMBO Rep*. 2021;22:e52013.
87. Ma X, Zhang Y, Gou D, Ma J, Du J, Wang C, Li S, Cui H. Metabolic reprogramming of microglia enhances proinflammatory cytokine release through EphA2/p38 MAPK pathway in Alzheimer's Disease. *J Alzheimers Dis*. 2022;88:771–85.
88. Baik SH, Kang S, Son SM, Mook-Jung I. Microglia contributes to plaque growth by cell death due to uptake of amyloid beta in the brain of Alzheimer's disease mouse model. *Glia*. 2016;64:2274–90.
89. McIntosh A, Mela V, Harty C, Minogue AM, Costello DA, Kerskens C, Lynch MA. Iron accumulation in microglia triggers a cascade of events that leads to altered metabolism and compromised function in APP/PS1 mice. *Brain Pathol*. 2019;29:606–21.
90. Guillot-Sestier MV, Araiz AR, Mela V, Gaban AS, O'Neill E, Joshi L, Chouchani ET, Mills EL, Lynch MA. Microglial metabolism is a pivotal factor in sexual dimorphism in Alzheimer's disease. *Commun Biol*. 2021;4:711.
91. Holland R, McIntosh AL, Finucane OM, Mela V, Rubio-Araiz A, Timmons G, McCarthy SA, Gun'ko YK, Lynch MA. Inflammatory microglia are glycolytic and iron retentive and typify the microglia in APP/PS1 mice. *Brain Behav Immun*. 2018;68:183–96.

92. Sangineto M, Ciarnelli M, Cassano T, Radesco A, Moola A, Bukke VN, Romano A, Villani R, Kanwal H, Capitanio N, et al. Metabolic reprogramming in inflammatory microglia indicates a potential way of targeting inflammation in Alzheimer's disease. *Redox Biol.* 2023;66:102846.
93. Sha X, Lin J, Wu K, Lu J, Yu Z. The TRPV1-PKM2-SREBP1 axis maintains microglial lipid homeostasis in Alzheimer's disease. *Cell Death Dis.* 2025;16:14.
94. Marino KM, Squirrel JM, Chacko JV, Watters JW, Eliceiri KW, Ulland TK. Metabolic response of microglia to amyloid deposition during Alzheimer's disease progression in a mouse model. *bioRxiv* 2023.
95. Stincone A, Prigione A, Cramer T, Wamelink MM, Campbell K, Cheung E, Olin-Sandoval V, Gruning NM, Kruger A, Tauqeer Alam M, et al. The return of metabolism: biochemistry and physiology of the pentose phosphate pathway. *Biol Rev Camb Philos Soc.* 2015;90:927–63.
96. TeSlaa T, Ralser M, Fan J, Rabinowitz JD. The pentose phosphate pathway in health and disease. *Nat Metab.* 2023;5:1275–89.
97. Gimeno-Bayon J, Lopez-Lopez A, Rodríguez MJ, Mahy N. Glucose pathways adaptation supports acquisition of activated microglia phenotype. *J Neurosci Res.* 2014;92:723–31.
98. Ju HQ, Lin JF, Tian T, Xie D, Xu RH. NADPH homeostasis in cancer: functions, mechanisms and therapeutic implications. *Signal Transduct Target Ther.* 2020;5:231.
99. Ying W. NAD<sup>+</sup>/NADH and NADP<sup>+</sup>/NADPH in cellular functions and cell death: regulation and biological consequences. *Antioxid Redox Signal.* 2008;10:179–206.
100. Tiwari M. Glucose 6 phosphatase dehydrogenase (G6PD) and neurodegenerative disorders: Mapping diagnostic and therapeutic opportunities. *Genes Dis.* 2017;4:196–203.
101. Stanton RC. Glucose-6-phosphate dehydrogenase, NADPH, and cell survival. *IUBMB Life.* 2012;64:362–9.
102. Tang BL. Neuroprotection by glucose-6-phosphate dehydrogenase and the pentose phosphate pathway. *J Cell Biochem.* 2019;120:14285–95.
103. Stepanov YV, Golovynska I, Zhang R, Golovynskyi S, Stepanova LI, Gorbach O, Dovbynchuk T, Garmanchuk LV, Ohulchanskyi TY, Qu J. Near-infrared light reduces beta-amyloid-stimulated microglial toxicity and enhances survival of neurons: mechanisms of light therapy for Alzheimer's disease. *Alzheimers Res Ther.* 2022;14:84.
104. Wickstead ES, Karim HA, Manuel RE, Biggs CS, Getting SJ, McArthur S. Reversal of beta-amyloid-induced microglial toxicity in vitro by activation of Fpr2/3. *Oxid Med Cell Longev.* 2020;2020:2139192.
105. Morganti C, Bonora M, Ito K. Metabolism and HSC fate: what NADPH is made for. *Trends Cell Biol* 2024.
106. Ferreira LMR, Li AM, Serafim TL, Sobral MC, Alpoim MC, Urbano AM. Intermediary metabolism: an intricate network at the crossroads of cell fate and function. *Biochim Biophys Acta Mol Basis Dis.* 2020;1866:165887.
107. Lee TY. Lactate: a multifunctional signaling molecule. *Yeungnam Univ J Med.* 2021;38:183–93.
108. Rabinowitz JD, Enderback S. Lactate: the ugly duckling of energy metabolism. *Nat Metab.* 2020;2:566–71.
109. Gray LR, Tompkins SC, Taylor EB. Regulation of pyruvate metabolism and human disease. *Cell Mol Life Sci.* 2014;71:2577–604.
110. Voloboueva LA, Emery JF, Sun X, Giffard RG. Inflammatory response of microglial BV-2 cells includes a glycolytic shift and is modulated by mitochondrial glucose-regulated protein 75/mortalin. *FEBS Lett.* 2013;587:756–62.
111. Andersson AK, Ronnback L, Hansson E. Lactate induces tumour necrosis factor- $\alpha$ , interleukin-6 and interleukin-1 $\beta$  release in microglial- and astroglial-enriched primary cultures. *J Neurochem.* 2005;93:1327–33.
112. Kong L, Wang Z, Liang X, Wang Y, Gao L, Ma C. Monocarboxylate transporter 1 promotes classical microglial activation and pro-inflammatory effect via 6-phosphofructo-2-kinase/fructose-2, 6-bisphosphatase 3. *J Neuroinflammation.* 2019;16:240.
113. Nicola R, Madar R, Okun E. HCARI1-Mediated L-lactate signaling suppresses microglial phagocytosis. *Neuromolecular Med.* 2022;24:399–404.
114. Pan RY, He L, Zhang J, Liu X, Liao Y, Gao J, Liao Y, Yan Y, Li Q, Zhou X, et al. Positive feedback regulation of microglial glucose metabolism by histone H4 lysine 12 lactylation in Alzheimer's disease. *Cell Metab.* 2022;34(634–648):e636.
115. Fan H, Yang F, Xiao Z, Luo H, Chen H, Chen Z, Liu Q, Xiao Y. Lactylation: novel epigenetic regulatory and therapeutic opportunities. *Am J Physiol Endocrinol Metab.* 2023;324:E330–8.
116. Kalsbeek MJ, Mulder L, Yi CX. Microglia energy metabolism in metabolic disorder. *Mol Cell Endocrinol.* 2016;438:27–35.
117. Wang L, Pavlou S, Du X, Bhuckory M, Xu H, Chen M. Glucose transporter 1 critically controls microglial activation through facilitating glycolysis. *Mol Neurodegener.* 2019;14:2.
118. Wang X, Wu Y, Tian Y, Hu H, Zhao Y, Xue B, Sun Z, Wei A, Xie F, Qian LJ. GLUT1-mediated microglial proinflammatory activation contributes to the development of stress-induced spatial learning and memory dysfunction in mice. *Cell Biosci.* 2024;14:48.
119. Dewanjee S, Chakraborty P, Bhattacharya H, Chacko L, Singh B, Chaudhary A, Javvaji K, Pradhan SR, Vallamkonda J, Dey A, et al. Altered glucose metabolism in Alzheimer's disease: role of mitochondrial dysfunction and oxidative stress. *Free Radic Biol Med.* 2022;193:134–57.
120. Hunt A, Schonknecht P, Henze M, Seidl U, Haberkorn U, Schroder J. Reduced cerebral glucose metabolism in patients at risk for Alzheimer's disease. *Psychiatry Res.* 2007;155:147–54.
121. Kyratata N, Emsley HCA, Sparasci O, Parkes LM, Dickie BR. A systematic review of glucose transport alterations in Alzheimer's disease. *Front Neurosci.* 2021;15:626636.
122. Do TM, Alata W, Dodacki A, Traversy MT, Chacun H, Pradier L, Schermann JM, Farinotti R, Calon F, Bourasset F. Altered cerebral vascular volumes and solute transport at the blood-brain barriers of two transgenic mouse models of Alzheimer's disease. *Neuropharmacology.* 2014;81:311–7.
123. Hooijmans CR, Graven C, Dederen PJ, Tanila H, van Groen T, Kiliaan AJ. Amyloid beta deposition is related to decreased glucose transporter-1 levels and hippocampal atrophy in brains of aged APP/PS1 mice. *Brain Res.* 2007;1181:93–103.
124. Kuznetsova E, Schliebs R. beta-Amyloid, cholinergic transmission, and cerebrovascular system – a developmental study in a mouse model of Alzheimer's disease. *Curr Pharm Des.* 2013;19:6749–65.
125. Mosconi L, Mistur R, Switalski R, Tsui WH, Glodzik L, Li Y, Pirraglia E, De Santi S, Reisberg B, Wisniewski T, de Leon MJ. FDG-PET changes in brain glucose metabolism from normal cognition to pathologically verified Alzheimer's disease. *Eur J Nucl Med Mol Imaging.* 2009;36:811–22.
126. McDonald TS, Lerskiatphanich T, Woodruff TM, McCombe PA, Lee JD. Potential mechanisms to modify impaired glucose metabolism in neurodegenerative disorders. *J Cereb Blood Flow Metab.* 2023;43:26–43.
127. Choi H, Choi Y, Lee EJ, Kim H, Lee Y, Kwon S, Hwang DW, Lee DS. Alzheimer's Disease neuroimaging I: hippocampal glucose uptake as a surrogate of metabolic change of microglia in Alzheimer's disease. *J Neuroinflammation.* 2021;18:190.
128. Hasel P, Aisenberg WH, Bennett FC, Liddelow SA. Molecular and metabolic heterogeneity of astrocytes and microglia. *Cell Metab.* 2023;35:555–70.
129. Wang Q, Duan L, Li X, Wang Y, Guo W, Guan F, Ma S. Glucose metabolism, neural cell senescence and Alzheimer's disease. *Int J Mol Sci.* 2022;23:4351.
130. Ardanaz CG, Ramirez MJ, Solas M. Brain metabolic alterations in Alzheimer's disease. *Int J Mol Sci.* 2022;23:3785.
131. Saito ER, Miller JB, Harari O, Cruchaga C, Mihindukulasuriya KA, Kauwe JSK, Bickman BT. Alzheimer's disease alters oligodendrocytic glycolytic and ketolytic gene expression. *Alzheimers Dement.* 2021;17:1474–86.
132. Izquierdo P, Jolivet RB, Attwell D, Madry C. Amyloid plaques and normal ageing have differential effects on microglial Ca(2+) activity in the mouse brain. *Pflugers Arch.* 2024;476:257–70.
133. Xiang X, Wind K, Wiedemann T, Blume T, Shi Y, Briel N, Beyer L, Biechele G, Eckenweber F, Zatzepin A, et al. Microglial activation states drive glucose uptake and FDG-PET alterations in neurodegenerative diseases. *Sci Transl Med.* 2021;13:eabe5640.
134. Iwasaki Y, Takeshima Y, Fujio K. Basic mechanism of immune system activation by mitochondria. *Immunol Med.* 2020;43:142–7.
135. Xu Y, Chen Y, Zhang X, Ma J, Liu Y, Cui L, Wang F. Glycolysis in innate immune cells contributes to autoimmunity. *Front Immunol.* 2022;13:920029.
136. Zhang H, Ma J, Tang K, Huang B. Beyond energy storage: roles of glycogen metabolism in health and disease. *FEBS J.* 2021;288:3772–83.
137. Roach PJ. Glycogen and its metabolism. *Curr Mol Med.* 2002;2:101–20.

138. Adeva-Andany MM, Gonzalez-Lucan M, Donapetry-Garcia C, Fernandez-Fernandez C, Ameneiros-Rodriguez E. Glycogen metabolism in humans. *BBA Clin*. 2016;5:85–100.
139. Wu L, Wong CP, Swanson RA. Methodological considerations for studies of brain glycogen. *J Neurosci Res*. 2019;97:914–22.
140. Schmid T, Brune B. Prostanoids and resolution of inflammation - beyond the lipid-mediator class switch. *Front Immunol*. 2021;12:714042.
141. Jiang J, Dingleline R. Prostaglandin receptor EP2 in the crosshairs of anti-inflammation, anti-cancer, and neuroprotection. *Trends Pharmacol Sci*. 2013;34:413–23.
142. Minhas PS, Latif-Hernandez A, McReynolds MR, Durairaj AS, Wang Q, Rubin A, Joshi AU, He JQ, Gauba E, Liu L, et al. Restoring metabolism of myeloid cells reverses cognitive decline in ageing. *Nature*. 2021;590:122–8.
143. Bisht K, Sharma KP, Lecours C, Sanchez MG, El Hajj H, Milior G, Olmos-Alonso A, Gomez-Nicola D, Luheshi G, Vallieres L, et al. Dark microglia: a new phenotype predominantly associated with pathological states. *Glia*. 2016;64:826–39.
144. St-Pierre MK, Carrier M, Gonzalez Ibanez F, Simoncicova E, Wallman MJ, Vallieres L, Parent M, Tremblay ME. Ultrastructural characterization of dark microglia during aging in a mouse model of Alzheimer's disease pathology and in human post-mortem brain samples. *J Neuroinflammation*. 2022;19:235.
145. St-Pierre MK, Simoncicova E, Bogi E, Tremblay ME. Shedding light on the dark side of the microglia. *ASN Neuro*. 2020;12:1759091420925335.
146. Bisht K, Sharma K, Lacoste B, Tremblay ME. Dark microglia: why are they dark? *Commun Integr Biol*. 2016;9:e1230575.
147. Grabert K, Michael T, Karavolos MH, Clohisey S, Baillie JK, Stevens MP, Freeman TC, Summers KM, McColl BW. Microglial brain region-dependent diversity and selective regional sensitivities to aging. *Nat Neurosci*. 2016;19:504–16.
148. Grassi S, Giussani P, Mauri L, Prioni S, Sonnino S, Prinetti A. Lipid rafts and neurodegeneration: structural and functional roles in physiologic aging and neurodegenerative diseases. *J Lipid Res*. 2020;61:636–54.
149. Miller YI, Navia-Pelaez JM, Corr M, Yaksh TL. Lipid rafts in glial cells: role in neuroinflammation and pain processing. *J Lipid Res*. 2020;61:655–66.
150. Ademowo OS, Dias HKI, Burton DGA, Griffiths HR. Lipid (per) oxidation in mitochondria: an emerging target in the ageing process? *Biogerontology*. 2017;18:859–79.
151. Li Y, Li Z, Yang M, Wang F, Zhang Y, Li R, Li Q, Gong Y, Wang B, Fan B, et al. Decoding the temporal and regional specification of microglia in the developing human brain. *Cell Stem Cell*. 2022;29(620–634):e626.
152. Tansley S, Uttam S, Urena Guzman A, Yaqubi M, Pacis A, Parisien M, Deamond H, Wong C, Rabau O, Brown N, et al. Single-cell RNA sequencing reveals time- and sex-specific responses of mouse spinal cord microglia to peripheral nerve injury and links ApoE to chronic pain. *Nat Commun*. 2022;13:843.
153. Tan YL, Yuan Y, Tian L. Microglial regional heterogeneity and its role in the brain. *Mol Psychiatry*. 2020;25:351–67.
154. Deczkowska A, Keren-Shaul H, Weiner A, Colonna M, Schwartz M, Amit I. Disease-associated microglia: a universal immune sensor of neurodegeneration. *Cell*. 2018;173:1073–81.
155. Foley P. Lipids in Alzheimer's disease: a century-old story. *Biochim Biophys Acta*. 2010;1801:750–3.
156. Kamboh MI, Demirci FY, Wang X, Minster RL, Carrasquillo MM, Pankratz VS, Younkin SG, Saykin AJ, Alzheimer's Disease Neuroimaging I, Jun G, et al. Genome-wide association study of Alzheimer's disease. *Transl Psychiatry* 2012, 2:e117.
157. Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, et al. A unique microglia type associated with restricting development of Alzheimer's disease. *Cell*. 2017;169(1276–1290):e1217.
158. Claes C, Danhash EP, Hasselmann J, Chadarevian JP, Shabestari SK, England WE, Lim TE, Hidalgo JLS, Spitale RC, Davtyan H, Blurton-Jones M. Plaque-associated human microglia accumulate lipid droplets in a chimeric model of Alzheimer's disease. *Mol Neurodegener*. 2021;16:50.
159. Krasemann S, Madore C, Cialic R, Baufeld C, Calcagno N, El Fatimy R, Beckers L, O'Loughlin E, Xu Y, Fanek Z, et al. The TREM2-APOE pathway drives the transcriptional phenotype of dysfunctional microglia in neurodegenerative diseases. *Immunity*. 2017;47(566–581):e569.
160. Sun N, Victor MB, Park YP, Xiong X, Scannail AN, Leary N, Prosper S, Viswanathan S, Luna X, Boix CA, et al. Human microglial state dynamics in Alzheimer's disease progression. *Cell*. 2023;186(4386–4403):e4329.
161. Wang Y, Cella M, Mallinson K, Ulrich JD, Young KL, Robinette ML, Gilfillan S, Krishnan GM, Sudhakar S, Zinselmeyer BH, et al. TREM2 lipid sensing sustains the microglial response in an Alzheimer's disease model. *Cell*. 2015;160:1061–71.
162. Leng L, Zhang J. Microglial metabolic reconfiguration provides a new strategy for the treatment of Alzheimer's disease. *Neural Regen Res*. 2023;18:1946–7.
163. Yen JJ, Yu II. The role of ApoE-mediated microglial lipid metabolism in brain aging and disease. *Immunometabolism (Cobham)*. 2023;5:e00018.
164. Mathys H, Davila-Velderrain J, Peng Z, Gao F, Mohammadi S, Young JZ, Menon M, He L, Abdurrob F, Jiang X, et al. Single-cell transcriptomic analysis of Alzheimer's disease. *Nature*. 2019;570:332–7.
165. Srinivasan K, Friedman BA, Etxeberria A, Huntley MA, van der Brug MP, Foreman O, Paw JS, Modrusan Z, Beach TG, Serrano GE, Hansen DV. Alzheimer's patient microglia exhibit enhanced aging and unique transcriptional activation. *Cell Rep*. 2020;31:107843.
166. Chen Y, Colonna M. Microglia in Alzheimer's disease at single-cell level. Are there common patterns in humans and mice? *J Exp Med*. 2021;218:e20202717.
167. Jung ES, Mook-Jung I. New microglia on the block. *Cell Metab*. 2020;31:664–6.
168. Mauerer R, Ebert S, Langmann T. High glucose, unsaturated and saturated fatty acids differentially regulate expression of ATP-binding cassette transporters ABCA1 and ABCG1 in human macrophages. *Exp Mol Med*. 2009;41:126–32.
169. Enkler L, Spang A. Functional interplay of lipid droplets and mitochondria. *FEBS Lett*. 2024;598:1235–51.
170. Fairley LH, Wong JH, Barron AM. Mitochondrial regulation of microglial immunometabolism in Alzheimer's disease. *Front Immunol*. 2021;12:624538.
171. Chandak PG, Radovic B, Aflaki E, Kolb D, Buchebner M, Frohlich E, Magnes C, Sinner F, Haemmerle G, Zechner R, et al. Efficient phagocytosis requires triacylglycerol hydrolysis by adipose triglyceride lipase. *J Biol Chem*. 2010;285:20192–201.
172. Yin B, Loike JD, Kako Y, Weinstock PH, Breslow JL, Silverstein SC, Goldberg IJ. Lipoprotein lipase regulates Fc receptor-mediated phagocytosis by macrophages maintained in glucose-deficient medium. *J Clin Invest*. 1997;100:649–57.
173. Zhang Y, Chen K, Sloan SA, Bennett ML, Scholze AR, O'Keefe S, Phatnani HP, Guarnieri P, Caneda C, Ruderisch N, et al. An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. *J Neurosci*. 2014;34:11929–47.
174. Flowers A, Bell-Temin H, Jalloh A, Stevens SM Jr, Bickford PC. Proteomic analysis of aged microglia: shifts in transcription, bioenergetics, and nutrient response. *J Neuroinflammation*. 2017;14:96.
175. Yin F. Lipid metabolism and Alzheimer's disease: clinical evidence, mechanistic link and therapeutic promise. *FEBS J*. 2023;290:1420–53.
176. Shippy DC, Ulland TK. Lipid metabolism transcriptomics of murine microglia in Alzheimer's disease and neuroinflammation. *Sci Rep*. 2023;13:14800.
177. K DB, Tang M, Reigan P, R HE. Genetic Variants of Lipoprotein Lipase and Regulatory Factors Associated with Alzheimer's Disease Risk. *Int J Mol Sci*. 2020;(21):8338.
178. Bruce KD, Gorkhali S, Given K, Coates AM, Boyle KE, Macklin WB, Eckel RH. Lipoprotein lipase is a feature of alternatively-activated microglia and may facilitate lipid uptake in the CNS during demyelination. *Front Mol Neurosci*. 2018;11:57.
179. Loving BA, Tang M, Neal MC, Gorkhali S, Murphy R, Eckel RH, Bruce KD. Lipoprotein lipase regulates microglial lipid droplet accumulation. *Cells*. 2021;10:198.
180. Martins IJ, Berger T, Sharman MJ, Verdile G, Fuller SJ, Martins RN. Cholesterol metabolism and transport in the pathogenesis of Alzheimer's disease. *J Neurochem*. 2009;111:1275–308.
181. Ma Y, Bao J, Zhao X, Shen H, Lv J, Ma S, Zhang X, Li Z, Wang S, Wang Q, Ji J. Activated cyclin-dependent kinase 5 promotes microglial phagocytosis of fibrillar beta-amyloid by up-regulating lipoprotein lipase expression. *Mol Cell Proteomics*. 2013;12:2833–44.

182. Dib S, Pahnke J, Gosselet F. Role of ABCA7 in human health and in Alzheimer's disease. *Int J Mol Sci*. 2021;22:4603.
183. Tanaka N, Abe-Dohmae S, Iwamoto N, Yokoyama S. Roles of ATP-binding cassette transporter A7 in cholesterol homeostasis and host defense system. *J Atheroscler Thromb*. 2011;18:274–81.
184. Satoh K, Abe-Dohmae S, Yokoyama S, St George-Hyslop P, Fraser PE. ATP-binding cassette transporter A7 (ABCA7) loss of function alters Alzheimer amyloid processing. *J Biol Chem*. 2015;290:24152–65.
185. Steinberg S, Stefansson H, Jonsson T, Johannsdottir H, Ingason A, Helgason H, Sulem P, Magnusson OT, Gudjonsson SA, Unnsteinsdottir U, et al. Loss-of-function variants in ABCA7 confer risk of Alzheimer's disease. *Nat Genet*. 2015;47:445–7.
186. De Roeck A, Van Broeckhoven C, Sleegers K. The role of ABCA7 in Alzheimer's disease: evidence from genomics, transcriptomics and methylomics. *Acta Neuropathol*. 2019;138:201–20.
187. Ma FC, Wang HF, Cao XP, Tan CC, Tan L, Yu JT. Meta-analysis of the association between variants in ABCA7 and Alzheimer's disease. *J Alzheimers Dis*. 2018;63:1261–7.
188. Aikawa T, Ren Y, Holm ML, Asmann YW, Alam A, Fitzgerald ML, Bu G, Kanekiyo T. ABCA7 regulates brain fatty acid metabolism during LPS-Induced acute inflammation. *Front Neurosci*. 2021;15:647974.
189. Dehghan A, Pinto RC, Karaman I, Huang J, Durainayagam BR, Ghanbari M, Nazeer A, Zhong Q, Liggi S, Whitley L, et al. Metabolome-wide association study on ABCA7 indicates a role of ceramide metabolism in Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2022;119:e2206083119.
190. Kim WS, Li H, Ruberu K, Chan S, Elliott DA, Low JK, Cheng D, Karl T, Garner B. Deletion of Abca7 increases cerebral amyloid-beta accumulation in the J20 mouse model of Alzheimer's disease. *J Neurosci*. 2013;33:4387–94.
191. Fu Y, Hsiao JH, Paxinos G, Halliday GM, Kim WS. ABCA7 mediates phagocytic clearance of amyloid-beta in the brain. *J Alzheimers Dis*. 2016;54:569–84.
192. Ma X, Prokopenko D, Wang N, Aikawa T, Choi Y, Zhang C, Lei D, Ren Y, Kawatani K, Starling SC, et al. Alzheimer's disease risk ABCA7 p.A696S variant disturbs the microglial response to amyloid pathology in mice. *Neurobiol Dis*. 2025;206:106813.
193. Wu X, Miller JA, Lee BTK, Wang Y, Ruedl C. Reducing microglial lipid load enhances beta amyloid phagocytosis in an Alzheimer's disease mouse model. *Sci Adv*. 2025;11:eadq6038.
194. Irshad Z, Dimitri F, Christian M, Zammit VA. Diacylglycerol acyltransferase 2 links glucose utilization to fatty acid oxidation in the brown adipocytes. *J Lipid Res*. 2017;58:15–30.
195. Walther TC, Chung J, Farese RV Jr. Lipid droplet biogenesis. *Annu Rev Cell Dev Biol*. 2017;33:491–510.
196. Flury A, Aljayousi L, Park HJ, Khakpour M, Mechler J, Aziz S, McGrath JD, Deme P, Sandberg C, Gonzalez Ibanez F, et al. A neurodegenerative cellular stress response linked to dark microglia and toxic lipid secretion. *Neuron*. 2024;113(4):554–71.
197. Lee J, Dimitry JM, Song JH, Son M, Sheehan PW, King MW, Travis Tabor G, Goo YA, Lazar MA, Petrucelli L, Musiek ES. Microglial REV-ERB $\alpha$  regulates inflammation and lipid droplet formation to drive tauopathy in male mice. *Nat Commun*. 2023;14:5197.
198. Li Y, Munoz-Mayorga D, Nie Y, Kang N, Tao Y, Lagerwall J, Pernaci C, Curtin G, Coufal NG, Mertens J, et al. Microglial lipid droplet accumulation in tauopathy brain is regulated by neuronal AMPK. *Cell Metab*. 2024;36(1351–1370):e1358.
199. Harayama T, Riezman H. Understanding the diversity of membrane lipid composition. *Nat Rev Mol Cell Biol*. 2018;19:281–96.
200. Shin JW, Lee JC. Roles of microglial membranes in Alzheimer's disease. *Curr Top Membr*. 2020;86:301–14.
201. Xu Y, Propson NE, Du S, Xiong W, Zheng H. Autophagy deficiency modulates microglial lipid homeostasis and aggravates tau pathology and spreading. *Proc Natl Acad Sci U S A*. 2021;118:e2023418118.
202. Ling ZN, Jiang YF, Ru JN, Lu JH, Ding B, Wu J. Amino acid metabolism in health and disease. *Signal Transduct Target Ther*. 2023;8:345.
203. Yoo HC, Yu YC, Sung Y, Han JM. Glutamine reliance in cell metabolism. *Exp Mol Med*. 2020;52:1496–516.
204. Suzumura A. Neurotoxicity by microglia: the mechanisms and potential therapeutic strategy. *Fukuoka Igaku Zasshi*. 2009;100:243–7.
205. Palmieri EM, Menga A, Lebrun A, Hooper DC, Butterfield DA, Mazzone M, Castegna A. Blockade of glutamine synthetase enhances inflammatory response in microglial cells. *Antioxid Redox Signal*. 2017;26:351–63.
206. Zhang Z, Li M, Li X, Feng Z, Luo G, Wang Y, Gao X. Glutamine metabolism modulates microglial NLRP3 inflammasome activity through mitophagy in Alzheimer's disease. *J Neuroinflammation*. 2024;21:261.
207. Ding L, Xu X, Li C, Wang Y, Xia X, Zheng JC. Glutaminase in microglia: a novel regulator of neuroinflammation. *Brain Behav Immun*. 2021;92:139–56.
208. Gao G, Zhao S, Xia X, Li C, Li C, Ji C, Sheng S, Tang Y, Zhu J, Wang Y, et al. Glutaminase C regulates microglial activation and pro-inflammatory exosome release: relevance to the pathogenesis of Alzheimer's disease. *Front Cell Neurosci*. 2019;13:264.
209. Cole GM, Ard MD. Influence of lipoproteins on microglial degradation of Alzheimer's amyloid beta-protein. *Microsc Res Tech*. 2000;50:316–24.
210. Jiang Q, Lee CY, Mandrekar S, Wilkinson B, Cramer P, Zelcer N, Mann K, Lamb B, Willson TM, Collins JL, et al. ApoE promotes the proteolytic degradation of A $\beta$ . *Neuron*. 2008;58:681–93.
211. Deng Y, Chen Q, Wan C, Sun Y, Huang F, Hu Y, Yang K. Microglia and macrophage metabolism: a regulator of cerebral gliomas. *Cell Biosci*. 2024;14:49.
212. Hu J, Baydyuk M, Huang JK. Impact of amino acids on microglial activation and CNS remyelination. *Curr Opin Pharmacol*. 2022;66:102287.
213. Schwarcz R, Bruno JP, Muchowski PJ, Wu HQ. Kynurenines in the mammalian brain: when physiology meets pathology. *Nat Rev Neurosci*. 2012;13:465–77.
214. Guillemin GJ, Brew BJ. Implications of the kynurenine pathway and quinolinic acid in Alzheimer's disease. *Redox Rep*. 2002;7:199–206.
215. Huang YS, Ogbuchi J, Clanchy FI, Williams RO, Stone TW. IDO and Kynurenine Metabolites in Peripheral and CNS Disorders. *Front Immunol*. 2020;11:388.
216. Savonije K, Weaver DF. The role of tryptophan metabolism in Alzheimer's disease. *Brain Sci*. 2023;13:292.
217. Lovelace MD, Varney B, Sundaram G, Lennon MJ, Lim CK, Jacobs K, Guillemin GJ, Brew BJ. Recent evidence for an expanded role of the kynurenine pathway of tryptophan metabolism in neurological diseases. *Neuropharmacology*. 2017;112:373–88.
218. Wu W, Nicolazzo JA, Wen L, Chung R, Stankovic R, Bao SS, Lim CK, Brew BJ, Cullen KM, Guillemin GJ. Expression of tryptophan 2,3-dioxygenase and production of kynurenine pathway metabolites in triple transgenic mice and human Alzheimer's disease brain. *PLoS ONE*. 2013;8:e59749.
219. Guillemin GJ, Brew BJ, Noonan CE, Takikawa O, Cullen KM. Indoleamine 2,3 dioxygenase and quinolinic acid immunoreactivity in Alzheimer's disease hippocampus. *Neuropathol Appl Neurobiol*. 2005;31:395–404.
220. Heyes MP, Saito K, Crowley JS, Davis LE, Demitrack MA, Der M, Dilling LA, Elia J, Kruesi MJ, Lackner A, et al. Quinolinic acid and kynurenine pathway metabolism in inflammatory and non-inflammatory neurological disease. *Brain*. 1992;115(Pt 5):1249–73.
221. Stone TW, Mackay GM, Forrest CM, Clark CJ, Darlington LG. Tryptophan metabolites and brain disorders. *Clin Chem Lab Med*. 2003;41:852–9.
222. Espey MG, Chernyshev ON, Reinhard JF Jr, Nambodiri MA, Colton CA. Activated human microglia produce the excitotoxin quinolinic acid. *NeuroReport*. 1997;8:431–4.
223. Heyes MP, Achim CL, Wiley CA, Major EO, Saito K, Markey SP. Human microglia convert l-tryptophan into the neurotoxin quinolinic acid. *Biochem J*. 1996;320(Pt 2):595–7.
224. Guillemin GJ. Quinolinic acid, the inescapable neurotoxin. *FEBS J*. 2012;279:1356–65.
225. Rahman A, Ting K, Cullen KM, Braid N, Brew BJ, Guillemin GJ. The excitotoxin quinolinic acid induces tau phosphorylation in human neurons. *PLoS ONE*. 2009;4:e6344.
226. Rath M, Muller I, Kropf P, Closs EI, Munder M. Metabolism via arginase or nitric oxide synthase: two competing arginine pathways in macrophages. *Front Immunol*. 2014;5:532.
227. Chhor V, Le Charpentier T, Lebon S, Ore MV, Celador IL, Jossereand J, Degos V, Jacotot E, Hagberg H, Savman K, et al. Characterization of phenotype markers and neuronotoxic potential of polarised primary microglia in vitro. *Brain Behav Immun*. 2013;32:70–85.
228. Cherry JD, Olschowka JA, O'Banion MK. Neuroinflammation and M2 microglia: the good, the bad, and the inflamed. *J Neuroinflammation*. 2014;11:98.
229. Jurga AM, Paleczna M, Kuter KZ. Overview of general and discriminating markers of differential microglia phenotypes. *Front Cell Neurosci*. 2020;14:198.

230. Brown GC. Nitric oxide and mitochondrial respiration. *Biochim Biophys Acta*. 1999;1411:351–69.
231. Caldeira C, Cunha C, Vaz AR, Falcao AS, Barateiro A, Seixas E, Fernandes A, Brites D. Key aging-associated alterations in primary microglia response to beta-amyloid stimulation. *Front Aging Neurosci*. 2017;9:277.
232. Combs CK, Karlo JC, Kao SC, Landreth GE. beta-Amyloid stimulation of microglia and monocytes results in TNFalpha-dependent expression of inducible nitric oxide synthase and neuronal apoptosis. *J Neurosci*. 2001;21:1179–88.
233. Saha RN, Pahan K. Regulation of inducible nitric oxide synthase gene in glial cells. *Antioxid Redox Signal*. 2006;8:929–47.
234. Saez TE, Pehar M, Vargas M, Barbeito L, Maccioni RB. Astrocytic nitric oxide triggers tau hyperphosphorylation in hippocampal neurons. *In Vivo*. 2004;18:275–80.
235. Cherry JD, Olschowka JA, O'Banion MK. Arginase 1+ microglia reduce Abeta plaque deposition during IL-1beta-dependent neuroinflammation. *J Neuroinflammation*. 2015;12:203.
236. Colonna M. The biology of TREM receptors. *Nat Rev Immunol*. 2023;23:580–94.
237. Wang S, Mustafa M, Yuede CM, Salazar SV, Kong P, Long H, Ward M, Siddiqui O, Paul R, Gilfillan S, et al. Anti-human TREM2 induces microglia proliferation and reduces pathology in an Alzheimer's disease model. *J Exp Med*. 2020;217:e20200785.
238. Takahashi K, Rochford CD, Neumann H. Clearance of apoptotic neurons without inflammation by microglial triggering receptor expressed on myeloid cells-2. *J Exp Med*. 2005;201:647–57.
239. Nugent AA, Lin K, van Lengerich B, Lianoglou S, Przybyla L, Davis SS, Llapashtica C, Wang J, Kim DJ, Xia D, et al. TREM2 regulates microglial cholesterol metabolism upon chronic phagocytic challenge. *Neuron*. 2020;105(837–854):e839.
240. Zhong L, Chen XF, Wang T, Wang Z, Liao C, Wang Z, Huang R, Wang D, Li X, Wu L, et al. Soluble TREM2 induces inflammatory responses and enhances microglial survival. *J Exp Med*. 2017;214:597–607.
241. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, Cruchaga C, Sassi C, Kauwe JS, Younkin S, et al. TREM2 variants in Alzheimer's disease. *N Engl J Med*. 2013;368:117–27.
242. Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, Bjornsson S, Huttenlocher J, Levey AI, Lah JJ, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med*. 2013;368:107–16.
243. Qin Q, Teng Z, Liu C, Li Q, Yin Y, Tang Y. TREM2, microglia, and Alzheimer's disease. *Mech Ageing Dev*. 2021;195:111438.
244. Wang L, Ouyang D, Li L, Cao Y, Wang Y, Gu N, Zhang Z, Li Z, Tang S, Tang H, et al. TREM2 affects DAM-like cell transformation in the acute phase of TBI in mice by regulating microglial glycolysis. *J Neuroinflammation*. 2025;22:6.
245. Lin M, Yu JX, Zhang WX, Lao FX, Huang HC. Roles of TREM2 in the pathological mechanism and the therapeutic strategies of Alzheimer's disease. *J Prev Alzheimers Dis*. 2024;11:1682–95.
246. Hou J, Chen Y, Grajales-Reyes G, Colonna M. TREM2 dependent and independent functions of microglia in Alzheimer's disease. *Mol Neurodegener*. 2022;17:84.
247. Ulland TK, Colonna M. TREM2 - a key player in microglial biology and Alzheimer disease. *Nat Rev Neurol*. 2018;14:667–75.
248. Feiten AF DK, van Lengerich B, Suh JH, Reifschneider A, Wefers B, Bartos LM, Wind-Mark K, Schlepckow K, Ulas T, De-Domenico E, Becker M, Khalin I, Davis SS, Wurst W, Plesnila N, Neher JJ, Brendel M, Lewcock JW, Di Paolo G, Capell A, Monroe KM, Schultze JL, Haass C. TREM2 expression level is critical for microglial state, metabolic capacity and efficacy of TREM2 agonism. *bioRxiv* 2024.
249. Park JC, Barahona-Torres N, Jang SY, Mok KY, Kim HJ, Han SH, Cho KH, Zhou X, Fu AKY, Ip NY, et al. Multi-Omics-based autophagy-related untypical subtypes in patients with cerebral amyloid pathology. *Adv Sci (Weinh)*. 2022;9:e2201212.
250. Piers TM, Cosker K, Mallach A, Johnson GT, Guerreiro R, Hardy J, Pocock JM. A locked immunometabolic switch underlies TREM2 R47H loss of function in human iPSC-derived microglia. *FASEB J*. 2020;34:2436–50.
251. Sayed FA, Kodama L, Fan L, Carling GK, Udeochu JC, Le D, Li Q, Zhou L, Wong MY, Horowitz R, et al. AD-linked R47H-TREM2 mutation induces disease-enhancing microglial states via AKT hyperactivation. *Sci Transl Med*. 2021;13:eabe3947.
252. Andreone BJ, Przybyla L, Llapashtica C, Rana A, Davis SS, van Lengerich B, Lin K, Shi J, Mei Y, Astarita G, et al. Alzheimer's-associated PLCgamma2 is a signaling node required for both TREM2 function and the inflammatory response in human microglia. *Nat Neurosci*. 2020;23:927–38.
253. Zhou Y, Song WM, Andhey PS, Swain A, Levy T, Miller KR, Poliani PL, Cominelli M, Grover S, Gilfillan S, et al. Human and mouse single-nucleus transcriptomics reveal TREM2-dependent and TREM2-independent cellular responses in Alzheimer's disease. *Nat Med*. 2020;26:131–42.
254. Sebastian Monasor L, Muller SA, Colombo AV, Tanriverog G, Konig J, Roth S, Liesz A, Berghofer A, Piechotta A, Prestel M, et al. Fibrillar Abeta triggers microglial proteome alterations and dysfunction in Alzheimer mouse models. *Elife*. 2020;9:e54083.
255. Henningfield CM, Arreola MA, Soni N, Spangenberg EE, Green KN. Microglia-specific ApoE knock-out does not alter Alzheimer's disease plaque pathogenesis or gene expression. *Glia*. 2022;70:287–302.
256. Yin Z, Rosenzweig N, Kleemann KL, Zhang X, Brandao W, Margeta MA, Schroeder C, Sivanathan KN, Silveira S, Gauthier C, et al. APOE4 impairs the microglial response in Alzheimer's disease by inducing TGFbeta-mediated checkpoints. *Nat Immunol*. 2023;24:1839–53.
257. Kontinen H, Cabral-da-Silva MEC, Ohtonen S, Wojciechowski S, Shakirzyanova A, Caligola S, Giugno R, Ishchenko Y, Hernandez D, Fazaludeen MF, et al. PSEN1DeltaE9, APPsw, and APOE4 confer disparate phenotypes in human iPSC-derived microglia. *Stem Cell Reports*. 2019;13:669–83.
258. Lin YT, Seo J, Gao F, Feldman HM, Wen HL, Penney J, Cam HP, Gjoneska E, Raja WK, Cheng J, et al. APOE4 causes widespread molecular and cellular alterations associated with Alzheimer's disease phenotypes in human iPSC-derived brain cell types. *Neuron*. 2018;98(1141–1154):e1147.
259. Millet A, Ledo JH, Tavazoie SF. An exhausted-like microglial population accumulates in aged and APOE4 genotype Alzheimer's brains. *Immunity*. 2024;57(153–170):e156.
260. Haney MS, Palovics R, Munson CN, Long C, Johansson PK, Yip O, Dong W, Rawat E, West E, Schlachetzki JCM, et al. APOE4/4 is linked to damaging lipid droplets in Alzheimer's disease microglia. *Nature*. 2024;628:154–61.
261. Murphy KB HD, Wolfs L, Mancuso R, De Strooper B, Marzi SJ: The APOE isoforms differentially shape the transcriptomic and epigenomic landscapes of human microglia in a xenotransplantation model of Alzheimer's disease. *bioRxiv* 2024.
262. Nguyen AT, Wang K, Hu G, Wang X, Miao Z, Azevedo JA, Suh E, Van Deerlin VM, Choi D, Roeder K, et al. APOE and TREM2 regulate amyloid-responsive microglia in Alzheimer's disease. *Acta Neuropathol*. 2020;140:477–93.
263. Yeh FL, Wang Y, Tom I, Gonzalez LC, Sheng M. TREM2 binds to apolipoproteins, including APOE and CLU/APOJ, and thereby facilitates uptake of amyloid-beta by microglia. *Neuron*. 2016;91:328–40.
264. Xue T, Ji J, Sun Y, Huang X, Cai Z, Yang J, Guo W, Guo R, Cheng H, Sun X. Sphingosine-1-phosphate, a novel TREM2 ligand, promotes microglial phagocytosis to protect against ischemic brain injury. *Acta Pharm Sin B*. 2022;12:1885–98.
265. Bedolla A, Wegman E, Weed M, Stevens MK, Ware K, Paranjpe A, Alkhimovitch A, Ifergan I, Taranov A, Peter JD, et al. Adult microglial TGFbeta1 is required for microglia homeostasis via an autocrine mechanism to maintain cognitive function in mice. *Nat Commun*. 2024;15:5306.
266. Wei W, Zhang L, Xin W, Pan Y, Tatenhorst L, Hao Z, Gerner ST, Huber S, Juenemann M, Butz M, et al. TREM2 regulates microglial lipid droplet formation and represses post-ischemic brain injury. *Biomed Pharmacother*. 2024;170:115962.
267. Atagi Y, Liu CC, Painter MM, Chen XF, Verbeeck C, Zheng H, Li X, Rademakers R, Kang SS, Xu H, et al. Apolipoprotein E is a ligand for triggering receptor expressed on myeloid cells 2 (TREM2). *J Biol Chem*. 2015;290:26043–50.
268. Jendresen C, Arskog V, Daws MR, Nilsson LN. The Alzheimer's disease risk factors apolipoprotein E and TREM2 are linked in a receptor signaling pathway. *J Neuroinflammation*. 2017;14:59.
269. Jay TR, von Saucken VE, Landreth GE. TREM2 in neurodegenerative diseases. *Mol Neurodegener*. 2017;12:56.



270. Wang Y, Lin Y, Wang L, Zhan H, Luo X, Zeng Y, Wu W, Zhang X, Wang F. TREM2 ameliorates neuroinflammatory response and cognitive impairment via PI3K/AKT/FoxO3a signaling pathway in Alzheimer's disease mice. *Aging* (Albany NY). 2020;12:20862–79.
271. Yuan P, Condello C, Keene CD, Wang Y, Bird TD, Paul SM, Luo W, Colonna M, Baddeley D, Grutzendler J. TREM2 haploinsufficiency in mice and humans impairs the microglia barrier function leading to decreased amyloid compaction and severe axonal dystrophy. *Neuron*. 2016;90:724–39.
272. Fitz NF, Nam KN, Wolfe CM, Letronne F, Playso BE, Iordanova BE, Kozai TDY, Biedrzycki RJ, Kagan VE, Tyurina YY, et al. Phospholipids of APOE lipoproteins activate microglia in an isoform-specific manner in pre-clinical models of Alzheimer's disease. *Nat Commun*. 2021;12:3416.
273. Park JC, Han JW, Lee W, Kim J, Lee SE, Lee D, Choi H, Han J, Kang YJ, Diep YN, et al. Microglia gravitate toward amyloid plaques surrounded by externalized phosphatidylserine via TREM2. *Adv Sci* (Weinh). 2024;11:e2400064.
274. Gratzke M, Schlachetzki JCM, D'Oliveira Albanus R, Jain N, Novotny B, Brase L, Rodriguez L, Mansel C, Kipnis M, O'Brien S, et al. TREM2-independent microgliosis promotes tau-mediated neurodegeneration in the presence of ApoE4. *Neuron*. 2023;111(202–219):e207.
275. Leyns CEG, Ulrich JD, Finn MB, Stewart FR, Koscal LJ, Remolina Serrano J, Robinson GO, Anderson E, Colonna M, Holtzman DM. TREM2 deficiency attenuates neuroinflammation and protects against neurodegeneration in a mouse model of tauopathy. *Proc Natl Acad Sci U S A*. 2017;114:11524–9.
276. Kober DL, Stuchell-Brereton MD, Klunder CE, Dean HB, Strickland MR, Steinberg DF, Nelson SS, Baban B, Holtzman DM, Frieden C, et al. Functional insights from biophysical study of TREM2 interactions with apoE and Abeta(1–42). *Alzheimers Dement* 2020.
277. Mai Z, Wei W, Yu H, Chen Y, Wang Y, Ding Y. Molecular recognition of the interaction between ApoE and the TREM2 protein. *Transl Neurosci*. 2022;13:93–103.
278. Li Z, Shue F, Zhao N, Shinohara M, Bu G. APOE2: protective mechanism and therapeutic implications for Alzheimer's disease. *Mol Neurodegener*. 2020;15:63.
279. Hu J, Liu CC, Chen XF, Zhang YW, Xu H, Bu G. Opposing effects of viral mediated brain expression of apolipoprotein E2 (apoE2) and apoE4 on apoE lipidation and Abeta metabolism in apoE4-targeted replacement mice. *Mol Neurodegener*. 2015;10:6.
280. Hanson AJ, Bayer-Carter JL, Green PS, Montine TJ, Wilkinson CW, Baker LD, Watson GS, Bonner LM, Callaghan M, Leverenz JB, et al. Effect of apolipoprotein E genotype and diet on apolipoprotein E lipidation and amyloid peptides: randomized clinical trial. *JAMA Neurol*. 2013;70:972–80.
281. Zhao J, Davis MD, Martens YA, Shinohara M, Graff-Radford NR, Yonkin SG, Wszolek ZK, Kanekiyo T, Bu G. APOE epsilon4/epsilon4 diminishes neurotrophic function of human iPSC-derived astrocytes. *Hum Mol Genet*. 2017;26:2690–700.
282. Lanfranco MF, Ng CA, Rebeck GW. ApoE lipidation as a therapeutic target in Alzheimer's disease. *Int J Mol Sci*. 2020;21:6336.
283. Guo JL, Braun D, Fitzgerald GA, Hsieh YT, Rouge L, Litvinchuk A, Steffek M, Propson NE, Heffner CM, Discenza C, et al. Decreased lipidated ApoE-receptor interactions confer protection against pathogenicity of ApoE and its lipid cargoes in lysosomes. *Cell*. 2025;188(187–206):e126.
284. Lindner K, Beckenbauer K, van Ek LC, Titeca K, de Leeuw SM, Awwad K, Hanke F, Korepanova AV, Rybin V, van der Kam EL, et al. Isoform- and cell-state-specific lipidation of ApoE in astrocytes. *Cell Rep*. 2022;38:110435.
285. Xia Z, Prescott EE, Urbanek A, Wareing HE, King MC, Olerinyova A, Dakin H, Leah T, Barnes KA, Matuszyk MM, et al. Co-aggregation with Apolipoprotein E modulates the function of Amyloid-beta in Alzheimer's disease. *Nat Commun*. 2024;15:4695.
286. Huynh TV, Davis AA, Ulrich JD, Holtzman DM. Apolipoprotein E and Alzheimer's disease: the influence of apolipoprotein E on amyloid-beta and other amyloidogenic proteins. *J Lipid Res*. 2017;58:824–36.
287. Wildsmith KR, Holley M, Savage JC, Skerrett R, Landreth GE. Evidence for impaired amyloid beta clearance in Alzheimer's disease. *Alzheimers Res Ther*. 2013;5:33.
288. Ibach M, Mathews M, Linnartz-Gerlach B, Theil S, Kumar S, Feederle R, Brustle O, Neumann H, Walter J. A reporter cell system for the triggering receptor expressed on myeloid cells 2 reveals differential effects of disease-associated variants on receptor signaling and activation by antibodies against the stalk region. *Glia*. 2021;69:1126–39.
289. Cosker K, Mallach A, Limaye J, Piers TM, Staddon J, Neame SJ, Hardy J, Pocock JM. Microglial signalling pathway deficits associated with the patient derived R47H TREM2 variants linked to AD indicate inability to activate inflammasome. *Sci Rep*. 2021;11:13316.
290. Kanekiyo T, Xu H, Bu G. ApoE and Abeta in Alzheimer's disease: accidental encounters or partners? *Neuron*. 2014;81:740–54.
291. Lessard CB, Malnik SL, Zhou Y, Ladd TB, Cruz PE, Ran Y, Mahan TE, Chakrabaty P, Holtzman DM, Ulrich JD, et al. High-affinity interactions and signal transduction between Abeta oligomers and TREM2. *EMBO Mol Med*. 2018;10(11):e9027.
292. Vander Heiden MG, DeBerardinis RJ. Understanding the Intersections between metabolism and cancer biology. *Cell*. 2017;168:657–69.
293. Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metab*. 2016;23:27–47.
294. Eltzschig HK, Carmeliet P. Hypoxia and inflammation. *N Engl J Med*. 2011;364:656–65.
295. Kierans SJ, Taylor CT. Regulation of glycolysis by the hypoxia-inducible factor (HIF): implications for cellular physiology. *J Physiol*. 2021;599:23–37.
296. Taylor CT, Scholz CC. The effect of HIF on metabolism and immunity. *Nat Rev Nephrol*. 2022;18:573–87.
297. Tannahill GM, Curtis AM, Adamik J, Palsson-McDermott EM, McGettrick AF, Goel G, Frezza C, Bernard NJ, Kelly B, Foley NH, et al. Succinate is an inflammatory signal that induces IL-1beta through HIF-1alpha. *Nature*. 2013;496:238–42.
298. McGettrick AF, O'Neill LAJ. The role of HIF in immunity and inflammation. *Cell Metab*. 2020;32:524–36.
299. Abdi Sarabi M, Shiri A, Aghapour M, Reichardt C, Brandt S, Mertens PR, Medunjanin S, Bruder D, Braun-Dullaeus RC, Weinert S. Normoxic HIF-1alpha stabilization caused by local inflammatory factors and its consequences in human coronary artery endothelial cells. *Cells*. 2022;11:3878.
300. Selak MA, Armour SM, MacKenzie ED, Boulahbel H, Watson DG, Mansfield KD, Pan Y, Simon MC, Thompson CB, Gottlieb E. Succinate links TCA cycle dysfunction to oncogenesis by inhibiting HIF-alpha prolyl hydroxylase. *Cancer Cell*. 2005;7:77–85.
301. Metzen E, Zhou J, Jelkmann W, Fandrey J, Brune B. Nitric oxide impairs normoxic degradation of HIF-1alpha by inhibition of prolyl hydroxylases. *Mol Biol Cell*. 2003;14:3470–81.
302. Li F, Sonveaux P, Rabbani ZN, Liu S, Yan B, Huang Q, Vujaskovic Z, Dewhirst MW, Li CY. Regulation of HIF-1alpha stability through S-nitrosylation. *Mol Cell*. 2007;26:63–74.
303. Braverman J, Stanley SA. Nitric oxide modulates macrophage responses to mycobacterium tuberculosis infection through activation of HIF-1alpha and repression of NF-kappaB. *J Immunol*. 2017;199:1805–16.
304. Palazon A, Goldrath AW, Nizet V, Johnson RS. HIF transcription factors, inflammation, and immunity. *Immunity*. 2014;41:518–28.
305. Yang F, Zhao D, Cheng M, Liu Y, Chen Z, Chang J, Dou Y. mTOR-mediated immunometabolic reprogramming nanomodulators enable sensitive switching of energy deprivation-induced microglial polarization for Alzheimer's disease management. *ACS Nano*. 2023;17:15724–41.
306. Liu P, Zhang T, Wu Y, Chen Q, Sun T, Jiang C. A peptide-drug conjugate-based nanoplateform for immunometabolic activation and in situ nerve regeneration in advanced-stage Alzheimer's disease. *Adv Mater*. 2024;36:e2408729.
307. Grubman A, Choo XY, Chew G, Ouyang JF, Sun G, Croft NP, Rossello FJ, Simmons R, Buckberry S, Landin DV, et al. Transcriptional signature in microglia associated with Abeta plaque phagocytosis. *Nat Commun*. 2021;12:3015.
308. March-Diaz R, Lara-Urena N, Romero-Molina C, Heras-Garvin A, Ortega-de San Luis C, Alvarez-Vergara MI, Sanchez-Garcia MA, Sanchez-Mejias E, Davila JC, Rosales-Nieves AE, et al. Hypoxia compromises the mitochondrial metabolism of Alzheimer's disease microglia via HIF1. *Nat Aging* 2021;1:385–399.
309. Gedam M, Comerota MM, Propson NE, Chen T, Jin F, Wang MC, Zheng H. Complement C3aR depletion reverses HIF-1alpha-induced metabolic impairment and enhances microglial response to Abeta pathology. *J Clin Invest*. 2023;133:e167501.
310. Herzig S, Shaw RJ. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol*. 2018;19:121–35.

311. Garcia D, Shaw RJ. AMPK: mechanisms of cellular energy sensing and restoration of metabolic balance. *Mol Cell*. 2017;66:789–800.
312. Lyons CL, Roche HM. Nutritional modulation of AMPK-impact upon metabolic-inflammation. *Int J Mol Sci*. 2018;19:3092.
313. Su R, Shao Y, Huang M, Liu D, Yu H, Qiu Y. Immunometabolism in cancer: basic mechanisms and new targeting strategy. *Cell Death Discovery*. 2024;10:236.
314. Lin M, Yu H, Xie Q, Xu Z, Shang P. Role of microglia autophagy and mitophagy in age-related neurodegenerative diseases. *Front Aging Neurosci*. 2022;14:1100133.
315. Wang S, Li H, Yuan M, Fan H, Cai Z. Role of AMPK in autophagy. *Front Physiol*. 2022;13:1015500.
316. Wang D, Wei Y, Tian J, He D, Zhang R, Ji X, Huang X, Sun J, Gao J, Wang Z, et al. Oxiracetam mediates neuroprotection through the regulation of microglia under hypoxia-ischemia neonatal brain injury in mice. *Mol Neurobiol*. 2021;58:3918–37.
317. Qing L, Fu J, Wu P, Zhou Z, Yu F, Tang J. Metformin induces the M2 macrophage polarization to accelerate the wound healing via regulating AMPK/mTOR/NLRP3 inflammasome singling pathway. *Am J Transl Res*. 2019;11:655–68.
318. Cui Y, Chen J, Zhang Z, Shi H, Sun W, Yi Q. The role of AMPK in macrophage metabolism, function and polarisation. *J Transl Med*. 2023;21:892.
319. Ou-Yang P, Cai ZY, Zhang ZH. Molecular regulation mechanism of microglial autophagy in the pathology of Alzheimer's disease. *Aging Dis*. 2023;14:1166–77.
320. Inoki K, Kim J, Guan KL. AMPK and mTOR in cellular energy homeostasis and drug targets. *Annu Rev Pharmacol Toxicol*. 2012;52:381–400.
321. Xu Y, Bai L, Yang X, Huang J, Wang J, Wu X, Shi J. Recent advances in anti-inflammation via AMPK activation. *Heliyon*. 2024;10:e33670.
322. Liang T, Zhang Y, Wu S, Chen Q, Wang L. The role of NLRP3 Inflammasome in Alzheimer's Disease and potential therapeutic targets. *Front Pharmacol*. 2022;13:845185.
323. Jung ES, Suh K, Han J, Kim H, Kang HS, Choi WS, Mook-Jung I. Amyloid- $\beta$  activates NLRP3 inflammasomes by affecting microglial immunometabolism through the Syk-AMPK pathway. *Aging Cell*. 2022;21:e13623.
324. Yu Q, Guo M, Zeng W, Zeng M, Zhang X, Zhang Y, Zhang W, Jiang X, Yu B. Interactions between NLRP3 inflammasome and glycolysis in macrophages: New insights into chronic inflammation pathogenesis. *Immun Inflamm Dis*. 2022;10:e581.
325. Olona A, Leishman S, Anand PK. The NLRP3 inflammasome: regulation by metabolic signals. *Trends Immunol*. 2022;43:978–89.
326. Ortega MA, De Leon-Oliva D, Garcia-Montero C, Fraile-Martinez O, Boaru DL, de Castro AV, Saez MA, Lopez-Gonzalez L, Bujan J, Alvarez-Mon MA, et al. Reframing the link between metabolism and NLRP3 inflammasome: therapeutic opportunities. *Front Immunol*. 2023;14:1232629.
327. Hawley SA, Fullerton MD, Ross FA, Schertzer JD, Chevtzoff C, Walker KJ, Pegg MW, Zibrova D, Green KA, Mustard KJ, et al. The ancient drug salicylate directly activates AMP-activated protein kinase. *Science*. 2012;336:918–22.
328. Yang F, Qin Y, Wang Y, Meng S, Xian H, Che H, Lv J, Li Y, Yu Y, Bai Y, Wang L. Metformin inhibits the NLRP3 inflammasome via AMPK/mTOR-dependent effects in diabetic cardiomyopathy. *Int J Biol Sci*. 2019;15:1010–9.
329. Jie F, Yang X, Yang B, Liu Y, Wu L, Lu B. Stigmasterol attenuates inflammatory response of microglia via NF- $\kappa$ B and NLRP3 signaling by AMPK activation. *Biomed Pharmacother*. 2022;153:113317.
330. Xu X, Gao W, Li L, Hao J, Yang B, Wang T, Li L, Bai X, Li F, Ren H, et al. Annexin A1 protects against cerebral ischemia-reperfusion injury by modulating microglia/macrophage polarization via FPR2/ALX-dependent AMPK-mTOR pathway. *J Neuroinflammation*. 2021;18:119.
331. Lambert JC, Ibrahim-Verbaas CA, Harold D, Naj AC, Sims R, Bellenguez C, DeStafano AL, Bis JC, Beecham GW, Grenier-Boley B, et al. Meta-analysis of 74,046 individuals identifies 11 new susceptibility loci for Alzheimer's disease. *Nat Genet*. 2013;45:1452–8.
332. Chou V, Pearce RV 2nd, Aylward AJ, Ashour N, Taga M, Terzioglu G, Fujita M, Fancher SB, Sigalov A, Benoit CR, et al. INPP5D regulates inflammasome activation in human microglia. *Nat Commun*. 2023;14:7552.
333. Cordero MD, Williams MR, Ryffel B. AMP-activated protein kinase regulation of the NLRP3 inflammasome during aging. *Trends Endocrinol Metab*. 2018;29:8–17.
334. Yun Q, Ma SF, Zhang WN, Gu M, Wang J. FoxG1 as a potential therapeutic target for Alzheimer's disease: modulating NLRP3 inflammasome via AMPK/mTOR autophagy pathway. *Cell Mol Neurobiol*. 2024;44:35.
335. Tsai AP, Lin PB, Dong C, Moutinho M, Casali BT, Liu Y, Lamb BT, Landreth GE, Oblak AL, Nho K. INPP5D expression is associated with risk for Alzheimer's disease and induced by plaque-associated microglia. *Neurobiol Dis*. 2021;153:105303.
336. Castranio EL, Hasel P, Haure-Mirande JV, Ramirez Jimenez AV, Hamilton BW, Kim RD, Glabe CG, Wang M, Zhang B, Gandy S, et al. Microglial INPP5D limits plaque formation and glial reactivity in the PSAPP mouse model of Alzheimer's disease. *Alzheimers Dement*. 2023;19:2239–52.
337. Terzioglu G, Young-Pearse TL. Microglial function, INPP5D/SHIP1 signaling, and NLRP3 inflammasome activation: implications for Alzheimer's disease. *Mol Neurodegener*. 2023;18:89.
338. Lin PB, Tsai AP, Soni D, Lee-Gosselin A, Moutinho M, Puntambekar SS, Landreth GE, Lamb BT, Oblak AL. INPP5D deficiency attenuates amyloid pathology in a mouse model of Alzheimer's disease. *Alzheimers Dement*. 2023;19:2528–37.
339. Yang L, Jiang Y, Shi L, Zhong D, Li Y, Li J, Jin R. AMPK: potential therapeutic target for Alzheimer's disease. *Curr Protein Pept Sci*. 2020;21:66–77.
340. Wang X, Zimmermann HR, Ma T. Therapeutic potential of AMP-Activated protein kinase in Alzheimer's disease. *J Alzheimers Dis*. 2019;68:33–8.
341. Zhao Z, Yan J, Huang L, Yang X. Phytochemicals targeting Alzheimer's disease via the AMP-activated protein kinase pathway, effects, and mechanisms of action. *Biomed Pharmacother*. 2024;173:116373.
342. Salminen A, Kaarniranta K, Haapasalo A, Soininen H, Hiltunen M. AMP-activated protein kinase: a potential player in Alzheimer's disease. *J Neurochem*. 2011;118:460–74.
343. Zhang N, Ji Q, Chen Y, Wen X, Shan F. TREM2 deficiency impairs the energy metabolism of Schwann cells and exacerbates peripheral neurological deficits. *Cell Death Dis*. 2024;15:193.
344. Saito M, Saito M, Das BC. Involvement of AMP-activated protein kinase in neuroinflammation and neurodegeneration in the adult and developing brain. *Int J Dev Neurosci*. 2019;77:48–59.
345. Baik SH, Ramanujan VK, Becker C, Fett S, Underhill DM, Wolf AJ. Hexokinase dissociation from mitochondria promotes oligomerization of VDAC that facilitates NLRP3 inflammasome assembly and activation. *Sci Immunol*. 2023;8:eade7652.
346. Guo D, Tong Y, Jiang X, Meng Y, Jiang H, Du L, Wu Q, Li S, Luo S, Li M, et al. Aerobic glycolysis promotes tumor immune evasion by hexokinase2-mediated phosphorylation of I $\kappa$ B $\alpha$ . *Cell Metab*. 2022;34:1312–1324.e1316.
347. Fairley LH, Lai KO, Wong JH, Chong WJ, Vincent AS, D'Agostino G, Wu X, Naik RR, Jayaraman A, Langley SR, et al. Mitochondrial control of microglial phagocytosis by the translocator protein and hexokinase 2 in Alzheimer's disease. *Proc Natl Acad Sci U S A*. 2023;120:e2209177120.
348. Guo D, Meng Y, Jiang X, Lu Z. Hexokinases in cancer and other pathologies. *Cell Insight*. 2023;2:100077.
349. Hu Y, Cao K, Wang F, Wu W, Mai W, Qiu L, Luo Y, Ge WP, Sun B, Shi L, et al. Dual roles of hexokinase 2 in shaping microglial function by gating glycolytic flux and mitochondrial activity. *Nat Metab*. 2022;4:1756–74.
350. Choi H, Mook-Jung I. Lipid fuel for hungry-angry microglia. *Nat Metab*. 2022;4:1223–4.
351. Tong Y, Liu X, Wu L, Xiang Y, Wang J, Cheng Y, Zhang C, Han B, Wang L, Yan D. Hexokinase 2 nonmetabolic function-mediated phosphorylation of I $\kappa$ B $\alpha$  enhances pancreatic ductal adenocarcinoma progression. *Cancer Sci*. 2024;115:2673–85.
352. Wang J, Shao F, Yang Y, Wang W, Yang X, Li R, Cheng H, Sun S, Feng X, Gao Y, et al. A non-metabolic function of hexokinase 2 in small cell lung cancer: promotes cancer cell stemness by increasing USP11-mediated CD133 stability. *Cancer Commun (Lond)*. 2022;42:1008–27.
353. Seki SM, Gaultier A. Exploring non-metabolic functions of glycolytic enzymes in immunity. *Front Immunol*. 2017;8:1549.
354. Stratoulis V, Venero JL, Tremblay ME, Joseph B. Microglial subtypes: diversity within the microglial community. *EMBO J*. 2019;38:e101997.
355. Prater KE, Green KJ, Mamde S, Sun W, Cochoit A, Smith CL, Chiou KL, Heath L, Rose SE, Wiley J, et al. Human microglia show unique transcriptional changes in Alzheimer's disease. *Nat Aging*. 2023;3:894–907.
356. Boche D, Gordon MN. Diversity of transcriptomic microglial phenotypes in aging and Alzheimer's disease. *Alzheimers Dement*. 2022;18:360–76.

357. Mathys H, Boix CA, Akay LA, Xia Z, Davila-Velderrain J, Ng AP, Jiang X, Abdelhady G, Galani K, Mantero J, et al. Single-cell multiregion dissection of Alzheimer's disease. *Nature*. 2024;632:858–68.
358. Nagata K, Hashimoto S, Joho D, Fujioka R, Matsuba Y, Sekiguchi M, Mihira N, Motooka D, Liu YC, Okuzaki D, et al: Tau Accumulation Induces Microglial State Alterations in Alzheimer's Disease Model Mice. *eNeuro*. 2024;11. PMID: 39592224.
359. Tyagi A, Mirita C, Taher N, Shah I, Moeller E, Tyagi A, Chong T, Pugazhenthi S. Metabolic syndrome exacerbates amyloid pathology in a comorbid Alzheimer's mouse model. *Biochim Biophys Acta Mol Basis Dis*. 2020;1866(10):165849.
360. Yuan C, Aierken A, Xie Z, Li N, Zhao J, Qing H. The age-related microglial transformation in Alzheimer's disease pathogenesis. *Neurobiol Aging*. 2020;92:82–91.
361. Ray WJ, Ashall F, Goate AM. Molecular pathogenesis of sporadic and familial forms of Alzheimer's disease. *Mol Med Today*. 1998;4:151–7.
362. Medina M, Khachaturian ZS, Rossor M, Avila J, Cedazo-Minguez A. Toward common mechanisms for risk factors in Alzheimer's syndrome. *Alzheimers Dement (N Y)*. 2017;3:571–8.
363. Lista S, O'Bryant SE, Blennow K, Dubois B, Hugon J, Zetterberg H, Hampel H. Biomarkers in Sporadic and Familial Alzheimer's Disease. *J Alzheimers Dis*. 2015;47:291–317.
364. de la Monte SM, Wands JR. Alzheimer's disease is type 3 diabetes-evidence reviewed. *J Diabetes Sci Technol*. 2008;2:1101–13.
365. Pasquier F, Boulogne A, Leys D, Fontaine P. Diabetes mellitus and dementia. *Diabetes Metab*. 2006;32:403–14.
366. Whitmer RA. Type 2 diabetes and risk of cognitive impairment and dementia. *Curr Neurol Neurosci Rep*. 2007;7:373–80.
367. Luchsinger JA, Reitz C, Patel B, Tang MX, Manly JJ, Mayeux R. Relation of diabetes to mild cognitive impairment. *Arch Neurol*. 2007;64:570–5.
368. Lloret A, Monllor P, Esteve D, Cervera-Ferri A, Lloret MA. Obesity as a risk factor for Alzheimer's disease: implication of leptin and glutamate. *Front Neurosci*. 2019;13:508.
369. Ebrahimpour S, Zakeri M, Esmaeili A. Crosstalk between obesity, diabetes, and alzheimer's disease: Introducing quercetin as an effective triple herbal medicine. *Ageing Res Rev*. 2020;62:101095.
370. Tian Y, Jing G, Ma M, Yin R, Zhang M. Microglial activation and polarization in type 2 diabetes-related cognitive impairment: a focused review of pathogenesis. *Neurosci Biobehav Rev*. 2024;165:105848.
371. Alexaki VI. The impact of obesity on microglial function: immune, metabolic and endocrine perspectives. *Cells*. 2021;10:1584.
372. Feng Z, Fang C, Ma Y, Chang J. Obesity-induced blood-brain barrier dysfunction: phenotypes and mechanisms. *J Neuroinflammation*. 2024;21:110.
373. Vargas-Soria M, Garcia-Alloza M, Corraliza-Gomez M. Effects of diabetes on microglial physiology: a systematic review of in vitro, preclinical and clinical studies. *J Neuroinflammation*. 2023;20:57.
374. Kim DK, Choi H, Lee W, Choi H, Hong SB, Jeong JH, Han J, Han JW, Ryu H, Kim JI, Mook-Jung I. Brain hypothyroidism silences the immune response of microglia in Alzheimer's disease animal model. *Sci Adv*. 1863;2024:eadi1863.

## Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.